

Vitrifying biological samples for cryo-imaging experiments

Gergely Papp

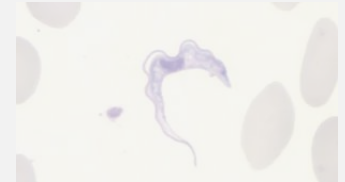
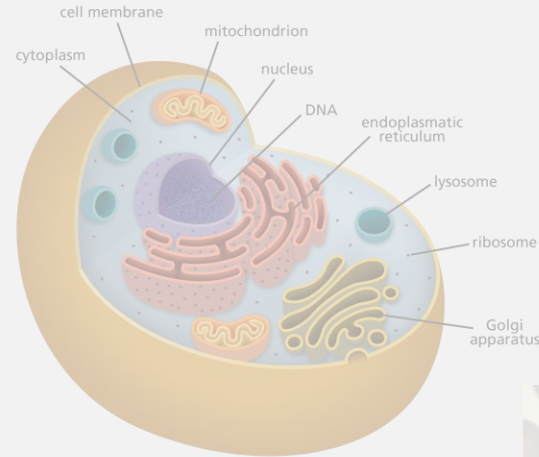
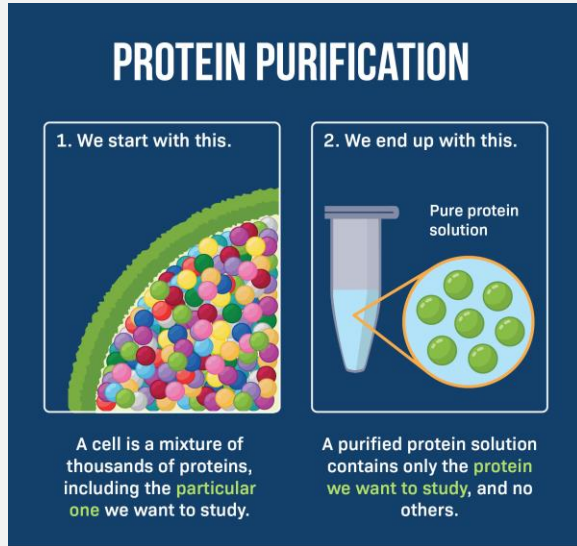
Team Leader

EIRO FORUM ESI – 14/5/2024

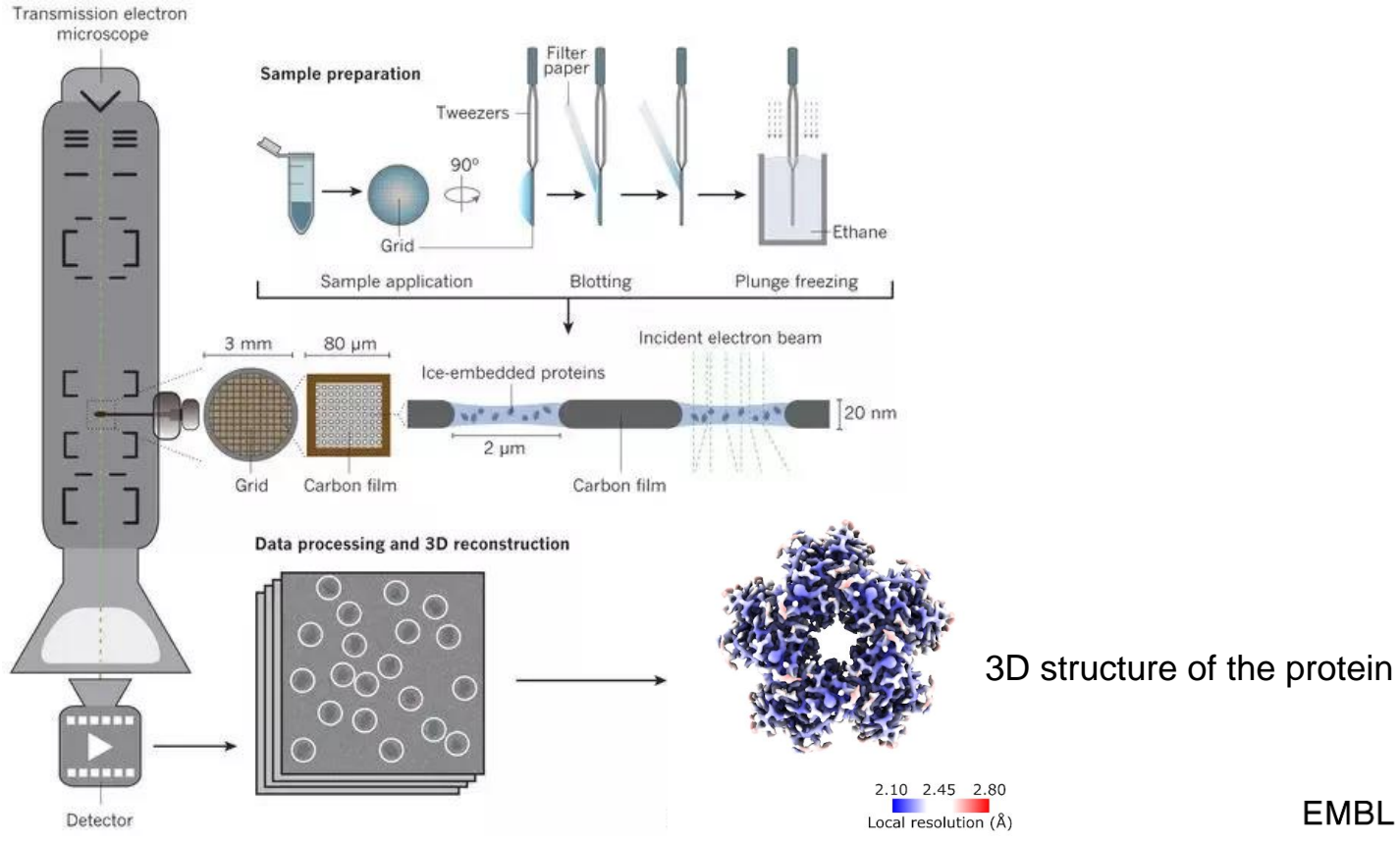


Which samples?

- Purified protein solutions (Single Particle Analysis)
 - Structure determination at atomic resolution
- Cells, monocellular organisms
 - In-situ study, proteins in their natural environment



Cryo-EM principle



The traditional sample preparation method

One session at the Vitrobot



Lengthy optimizations...



- Poor reproducibility of thin ice film production
 - Slow grid screening process
 - Multi-parametric optimizations
 - “No size fits all”

EasyGrid, automated Cryo-EM sample preparation



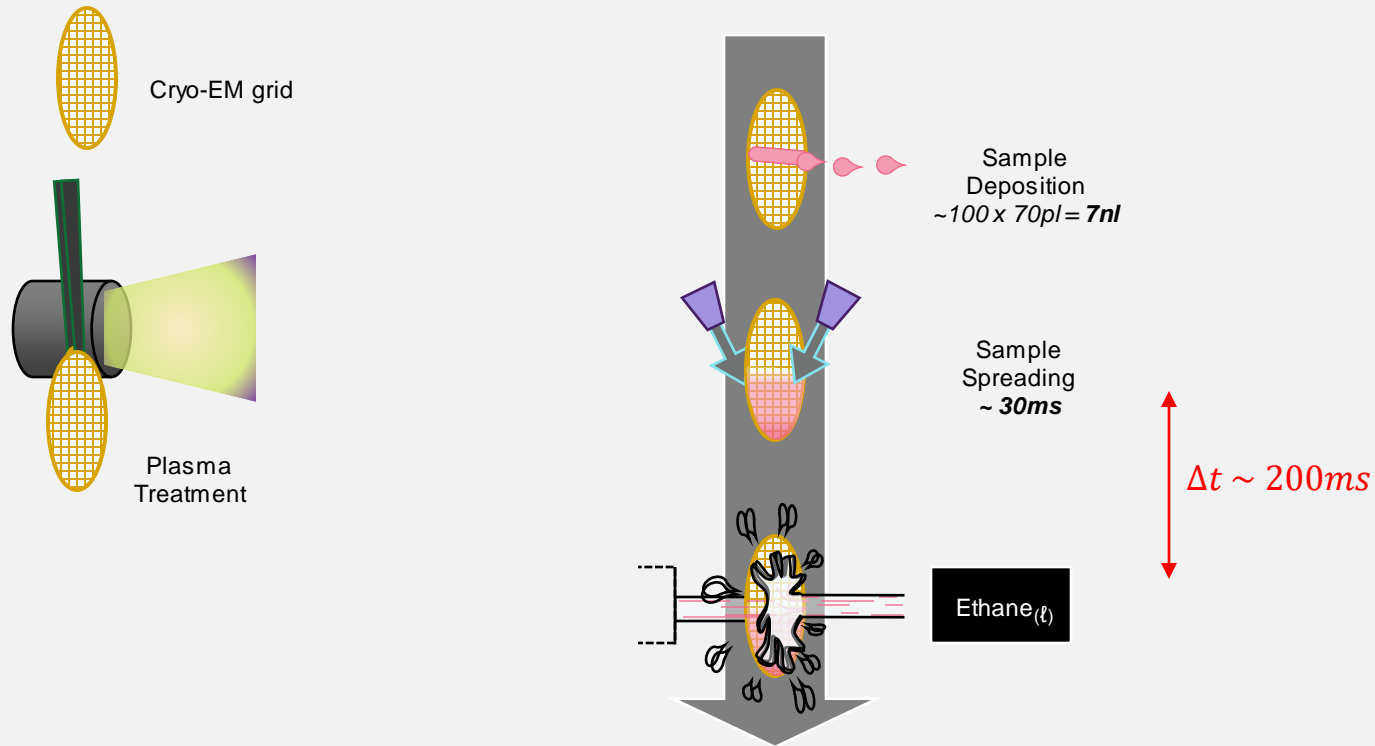
EMBL - GR



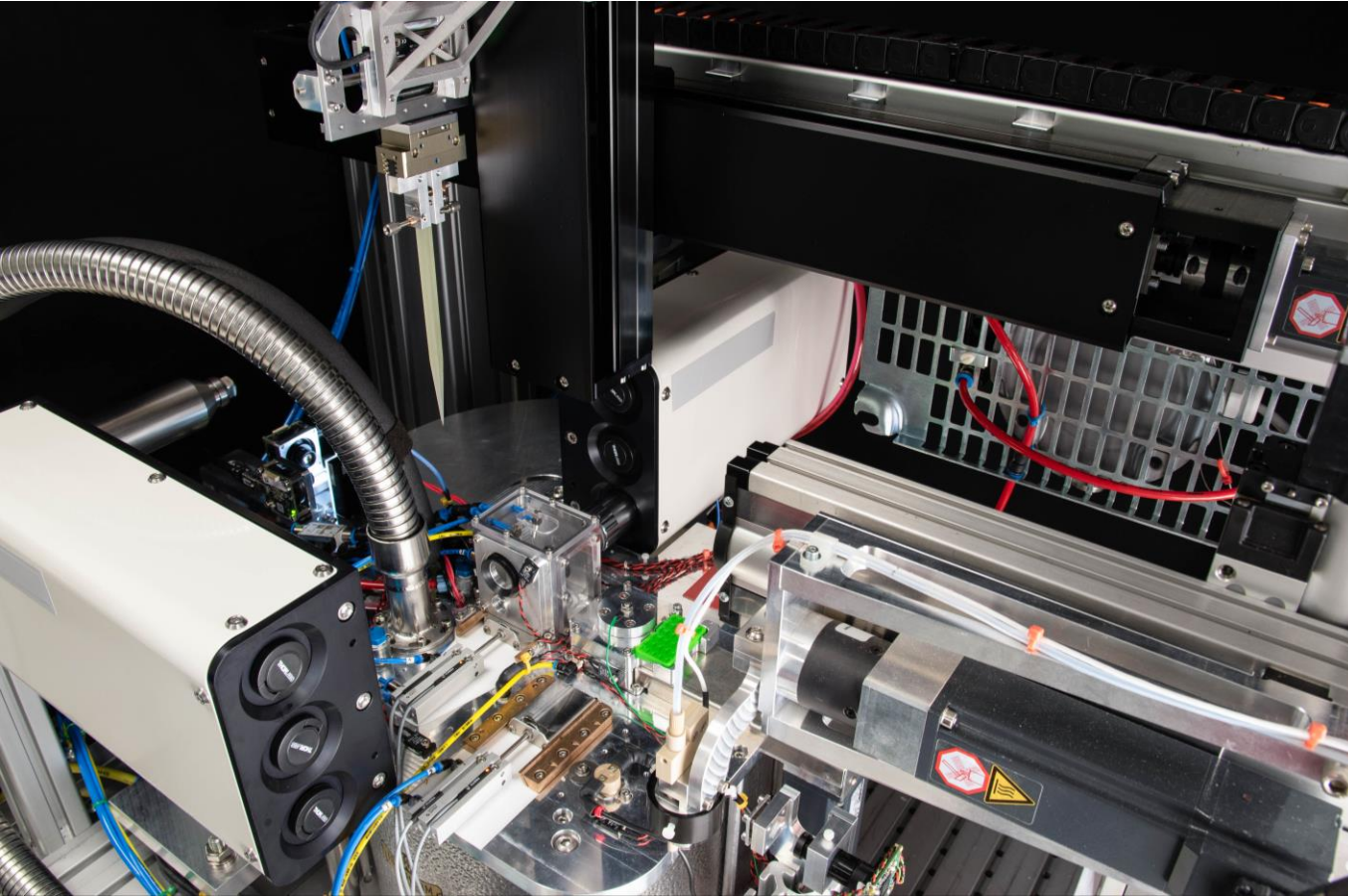
EMBL – HD
&
Imaging Centre



Preparation process for SPA samples

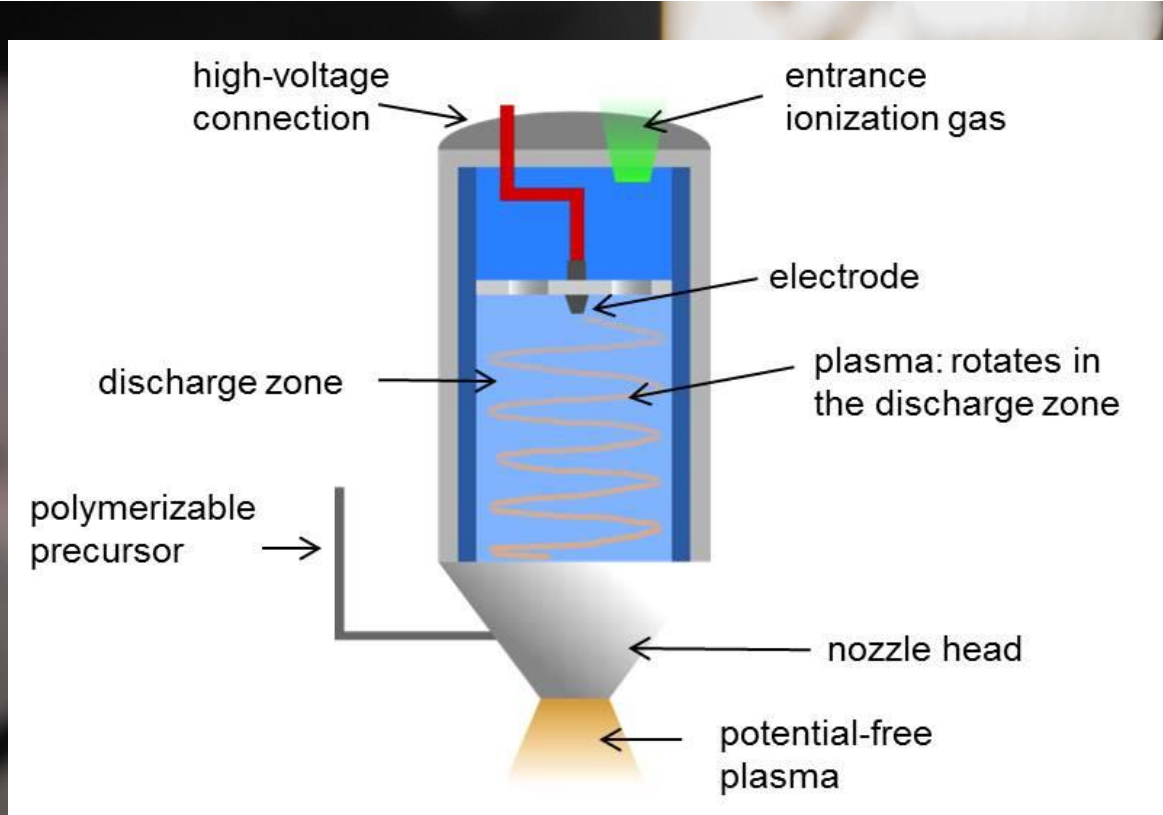


Components



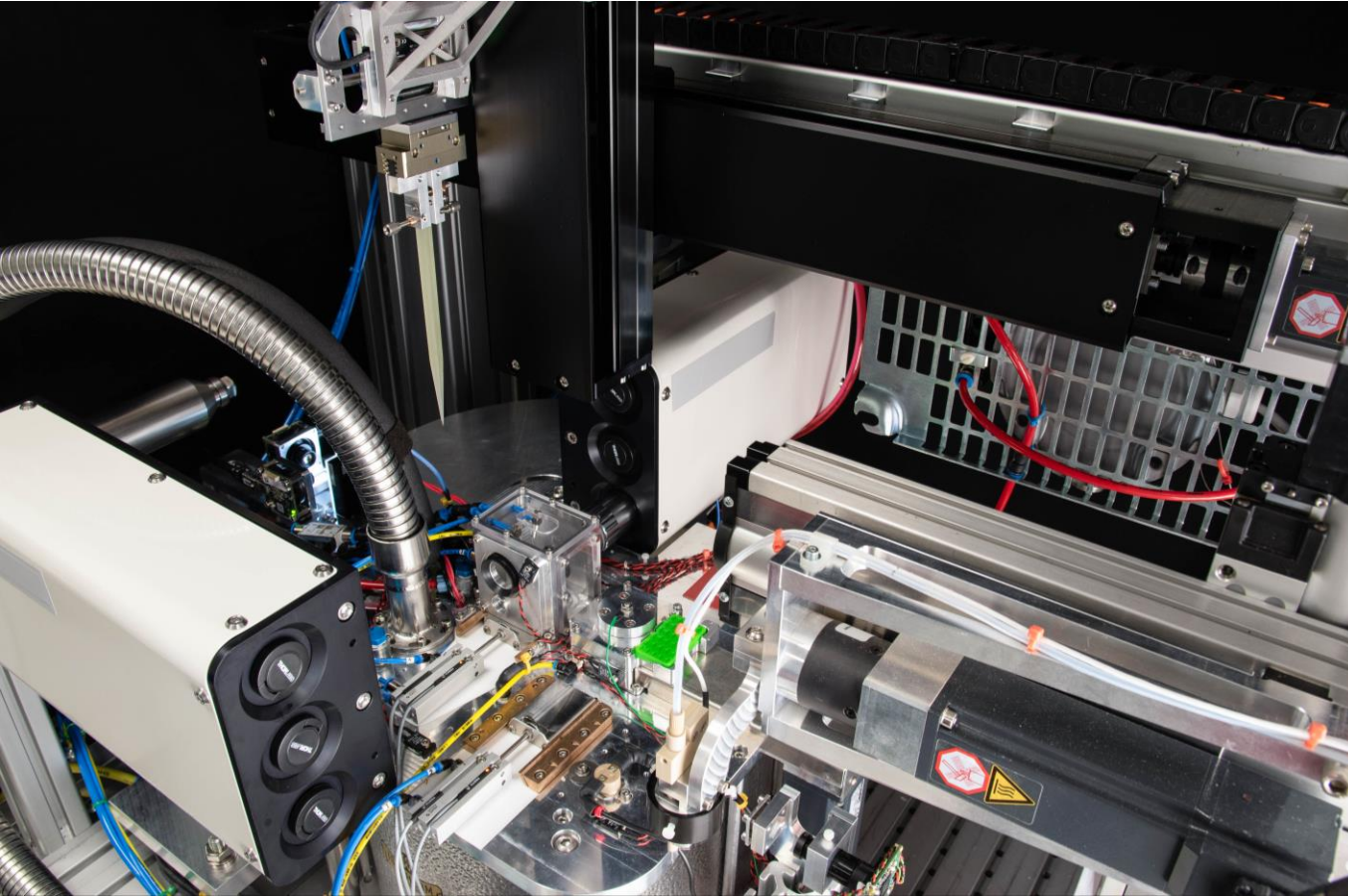
- Atmospheric plasma treatment

Components



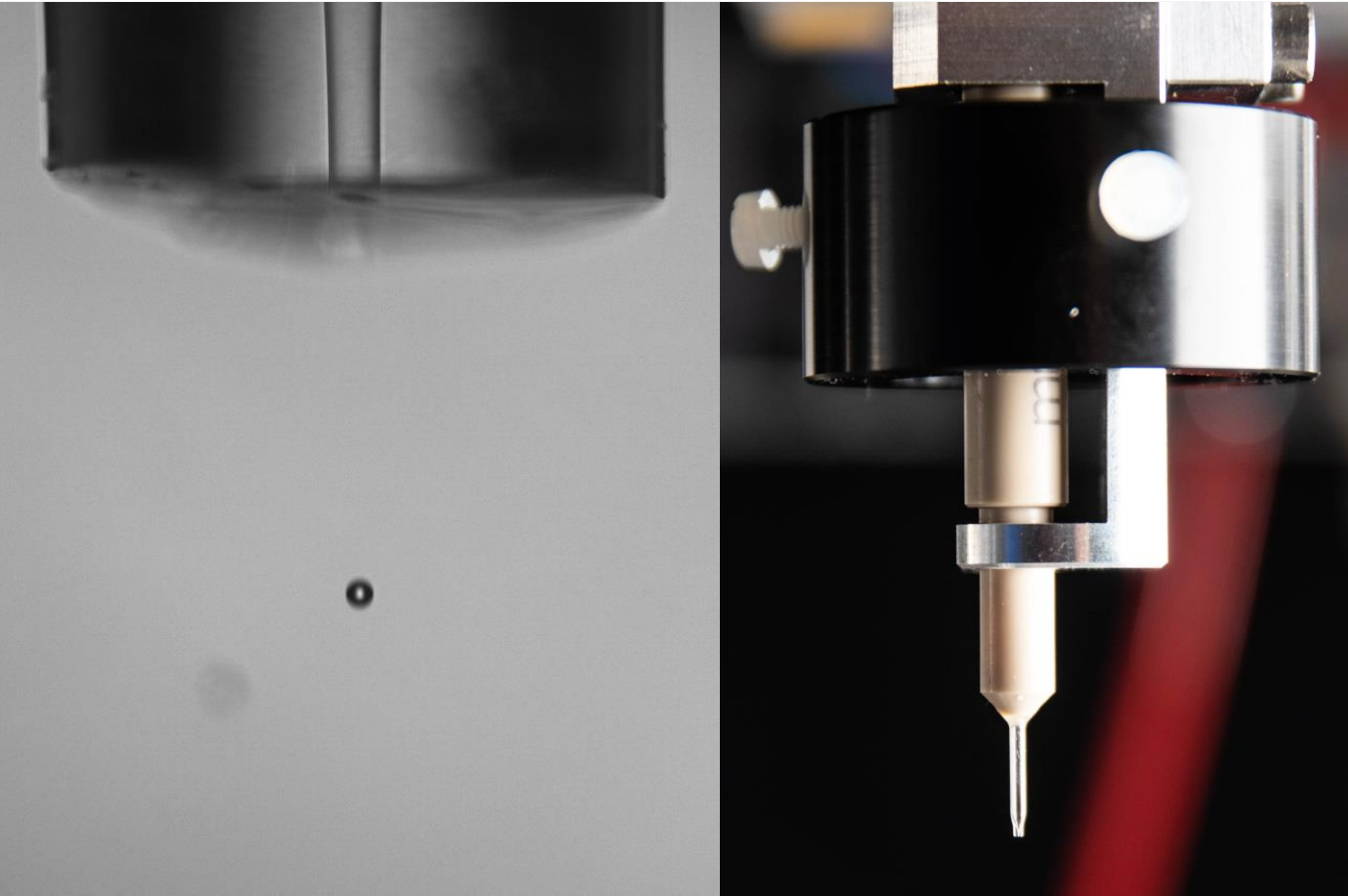
- Atmospheric plasma treatment
- **Advantage:**
 - In-line hydrophilic treatment
 - Minimum time between treatment and sample deposition
 - Possibility of material deposition and use of other gases

Components

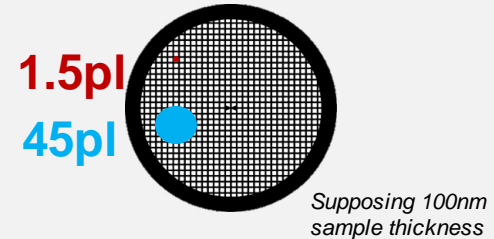


- Atmospheric plasma treatment
- Picoliter drop dispenser

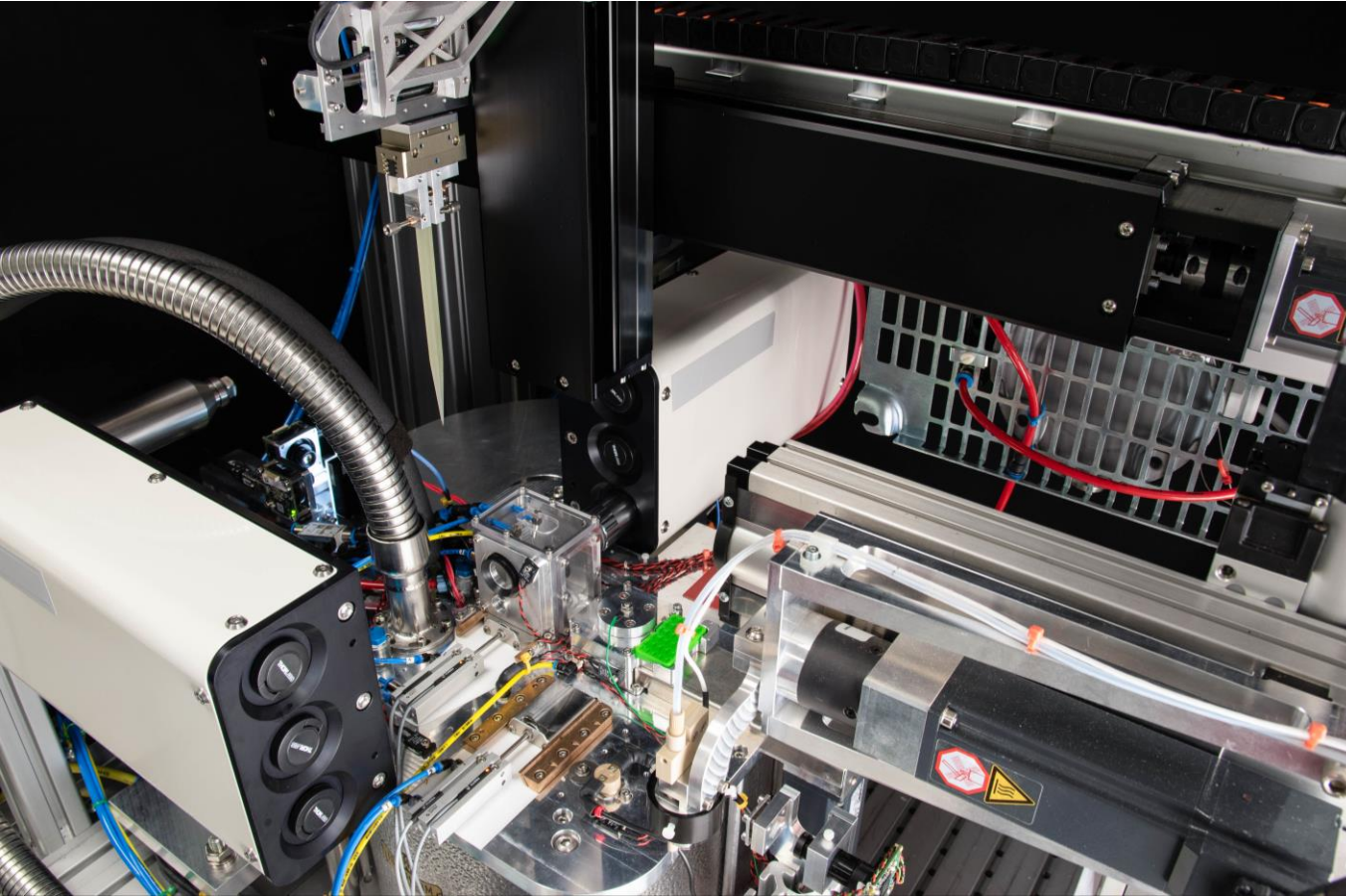
Components



- Atmospheric plasma treatment
- Picoliter drop dispenser
- **Advantage:**
 - Highly repeatable
 - Controlled deposition

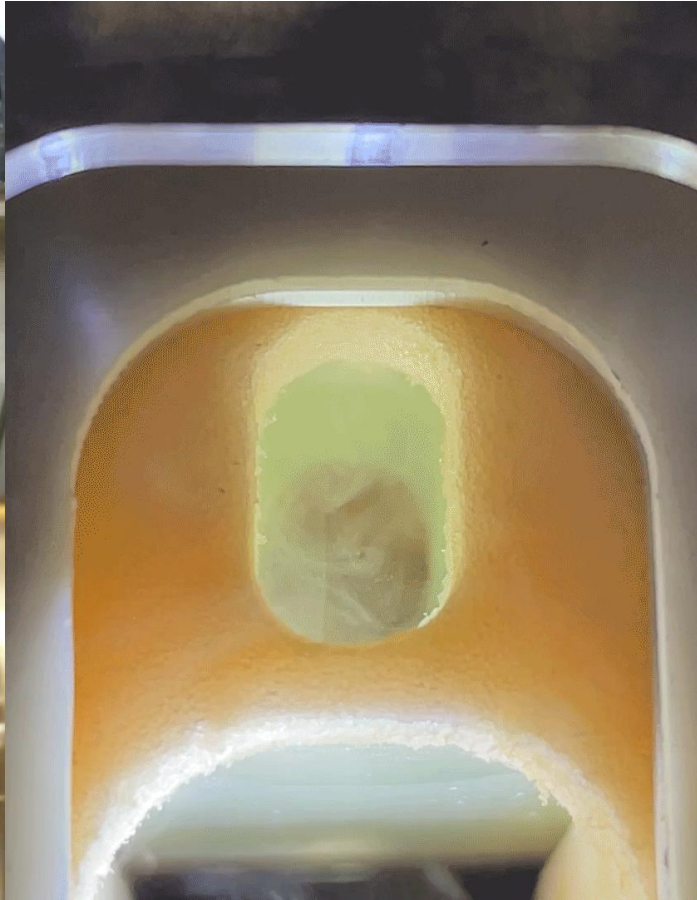
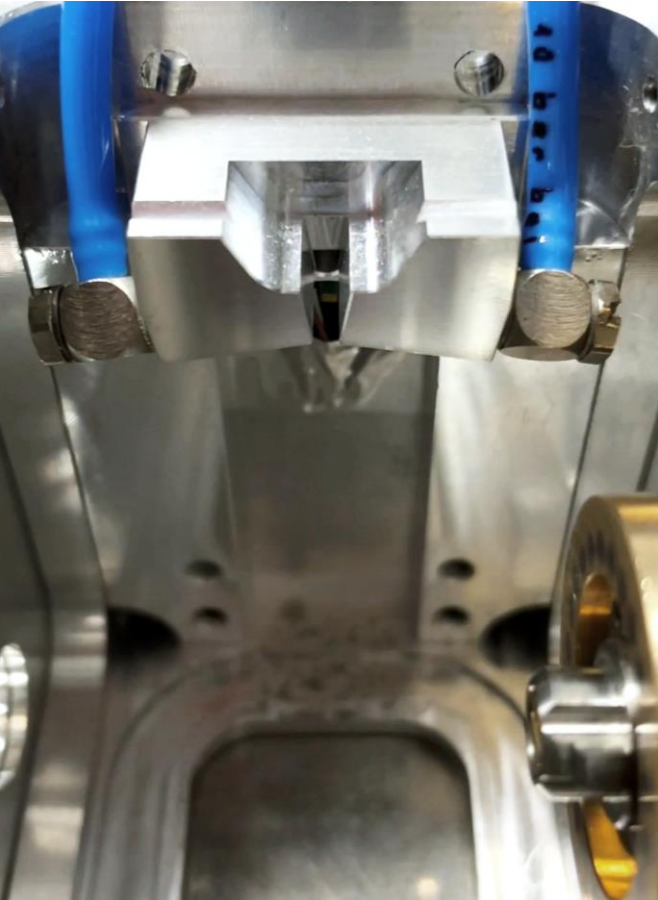


Components



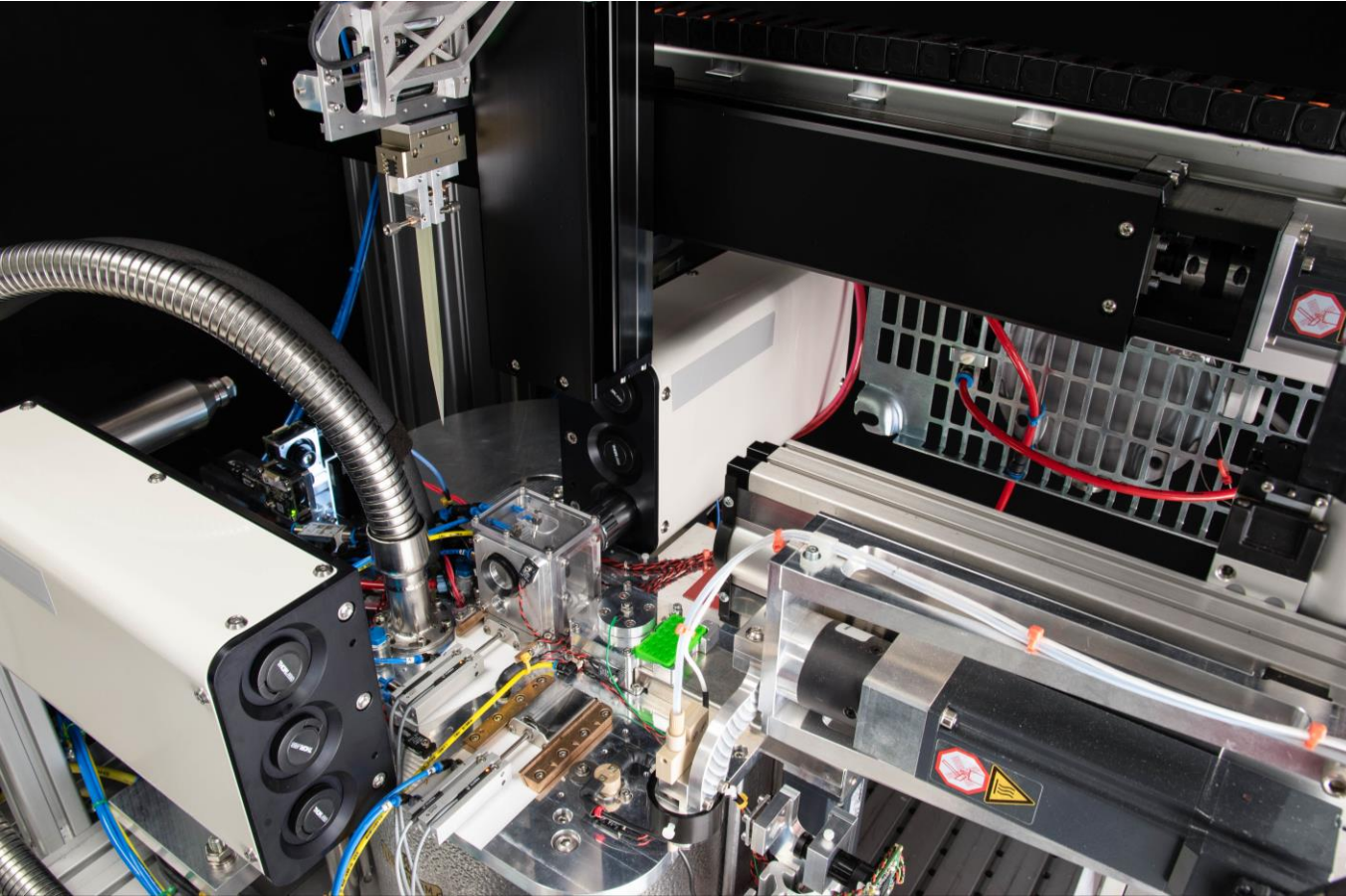
- Atmospheric plasma treatment
- Picoliter drop dispenser
- Chamber
 - Pressure wave generator
 - Humidity control
 - Ethane jet for vitrification

Components



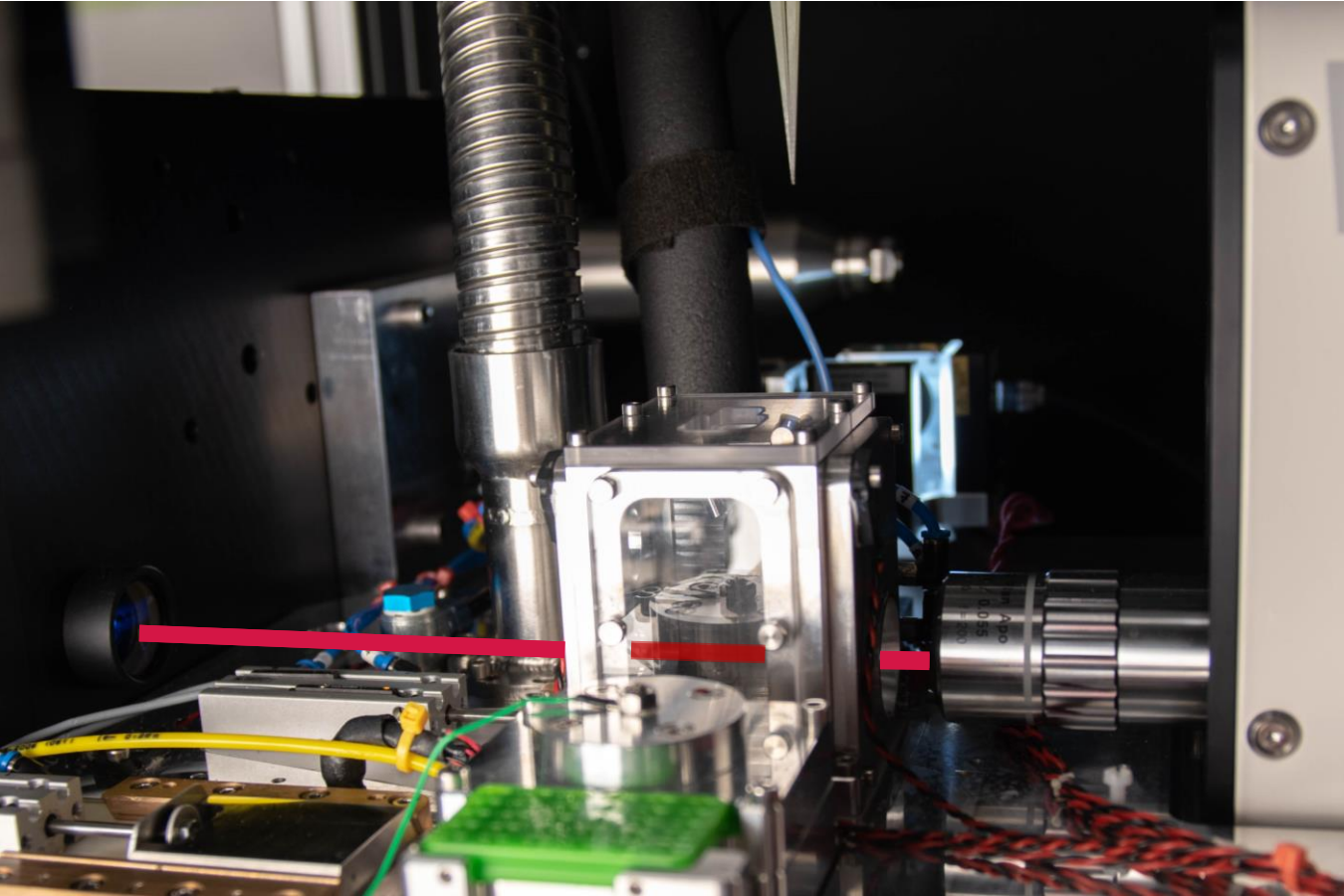
- Atmospheric plasma treatment
- Picoliter drop dispenser
- Chamber
 - Pressure wave generator
 - Humidity & temperature control
 - Ethane jet for vitrification
- **Advantage:**
 - Contactless spreading
 - Higher cooling rate

Components



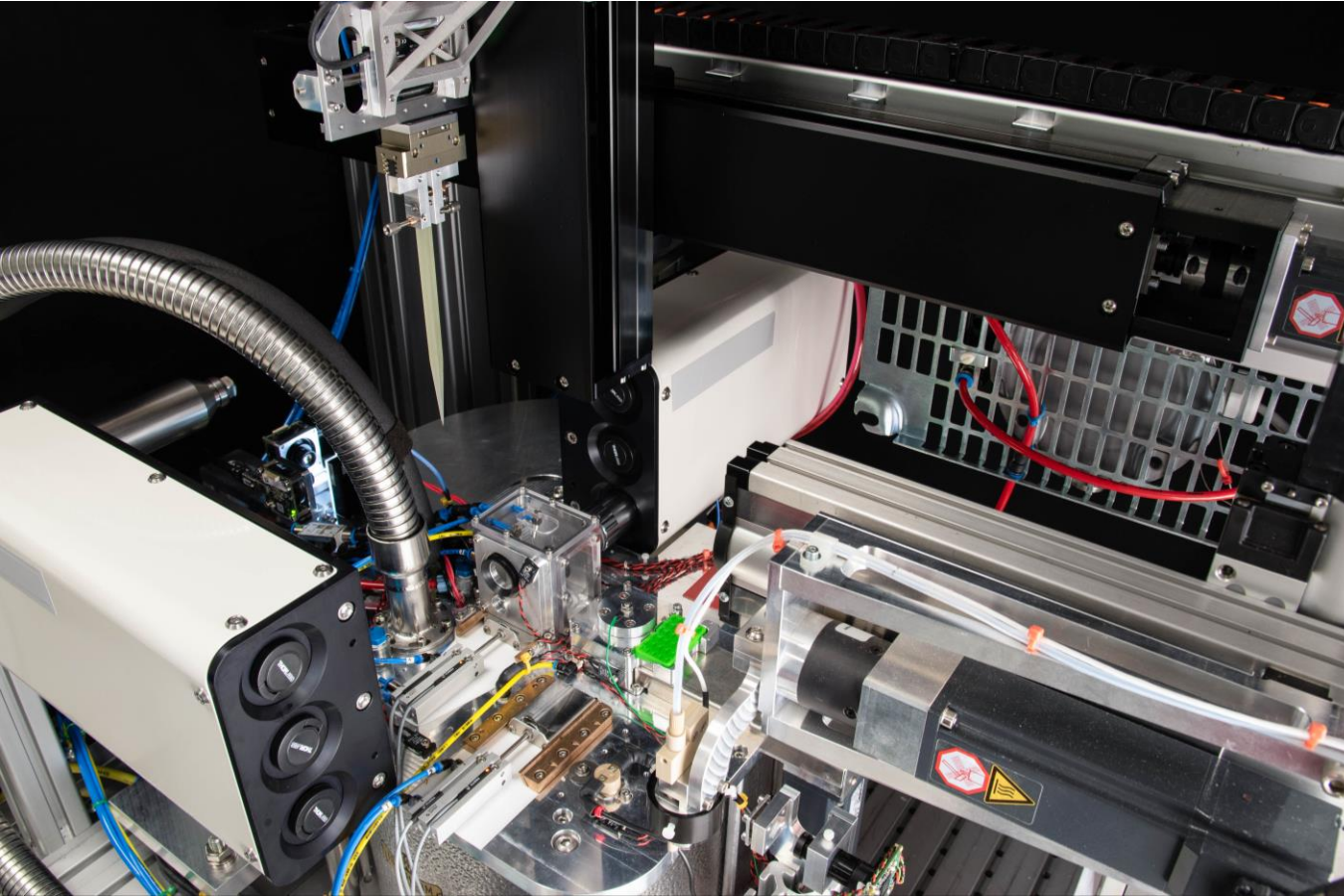
- Atmospheric plasma treatment
- Picoliter drop dispenser
- Chamber
 - Air blades (spreading)
 - Humidity control
 - Ethane jet for vitrification
- Digital Holographic Microscope (DHM)

Components



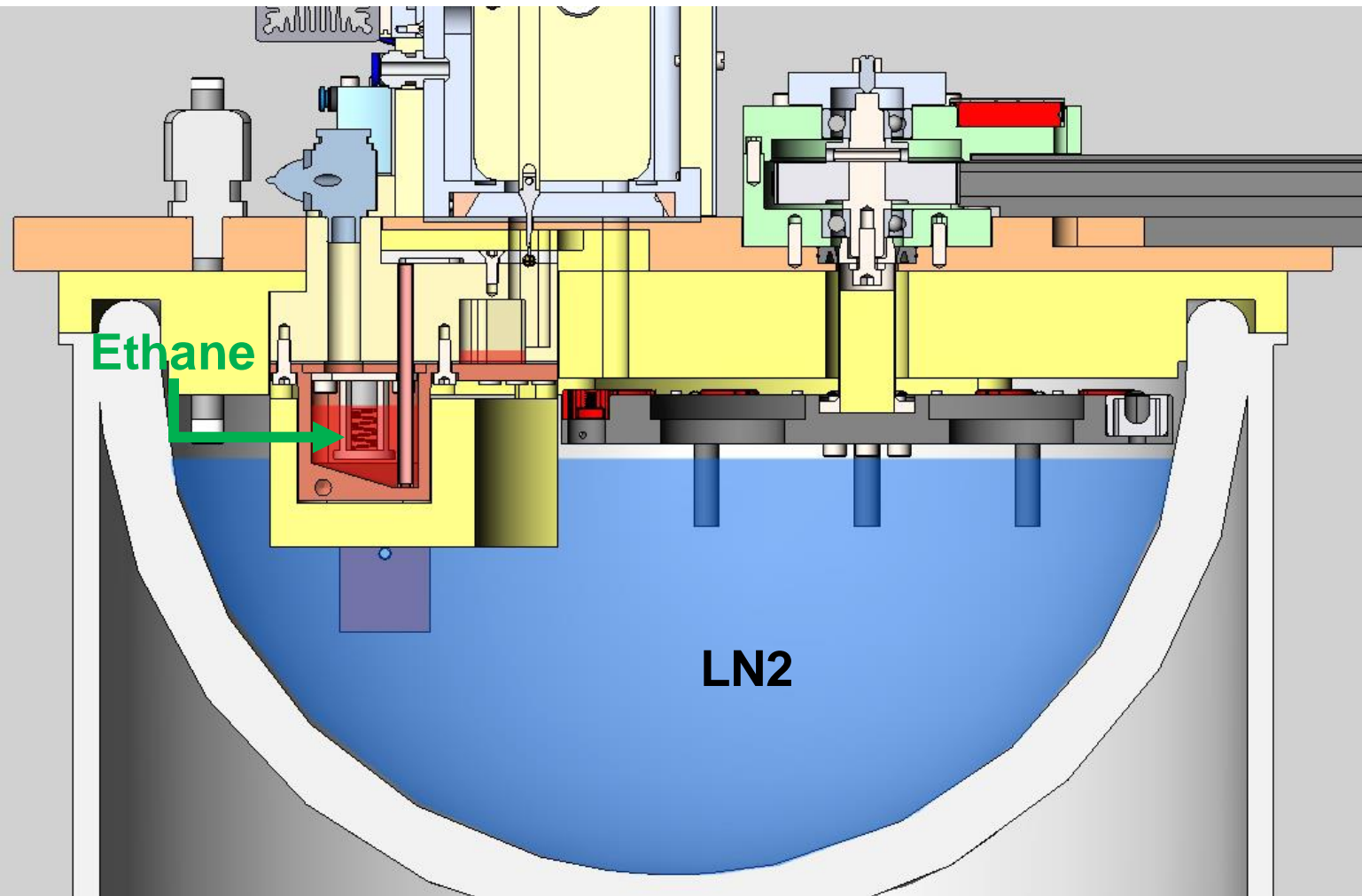
- Atmospheric plasma treatment
- Picoliter drop dispenser
- Chamber
 - Air blades (spreading)
 - Humidity control
 - Ethane jet for vitrification
- Digital Holographic Microscope (DHM)
 - Control of the sample thickness

Components



- Atmospheric plasma treatment
- Picoliter drop dispenser
- Chamber
 - Air blades (spreading)
 - Humidity control
 - Ethane jet for vitrification
- Digital Holographic Microscope (DHM)
- Dewar
 - Storage in Liquid Nitrogen
 - Ethane reservoir

C



c plasma

op dispenser

(spreading)

ontrol

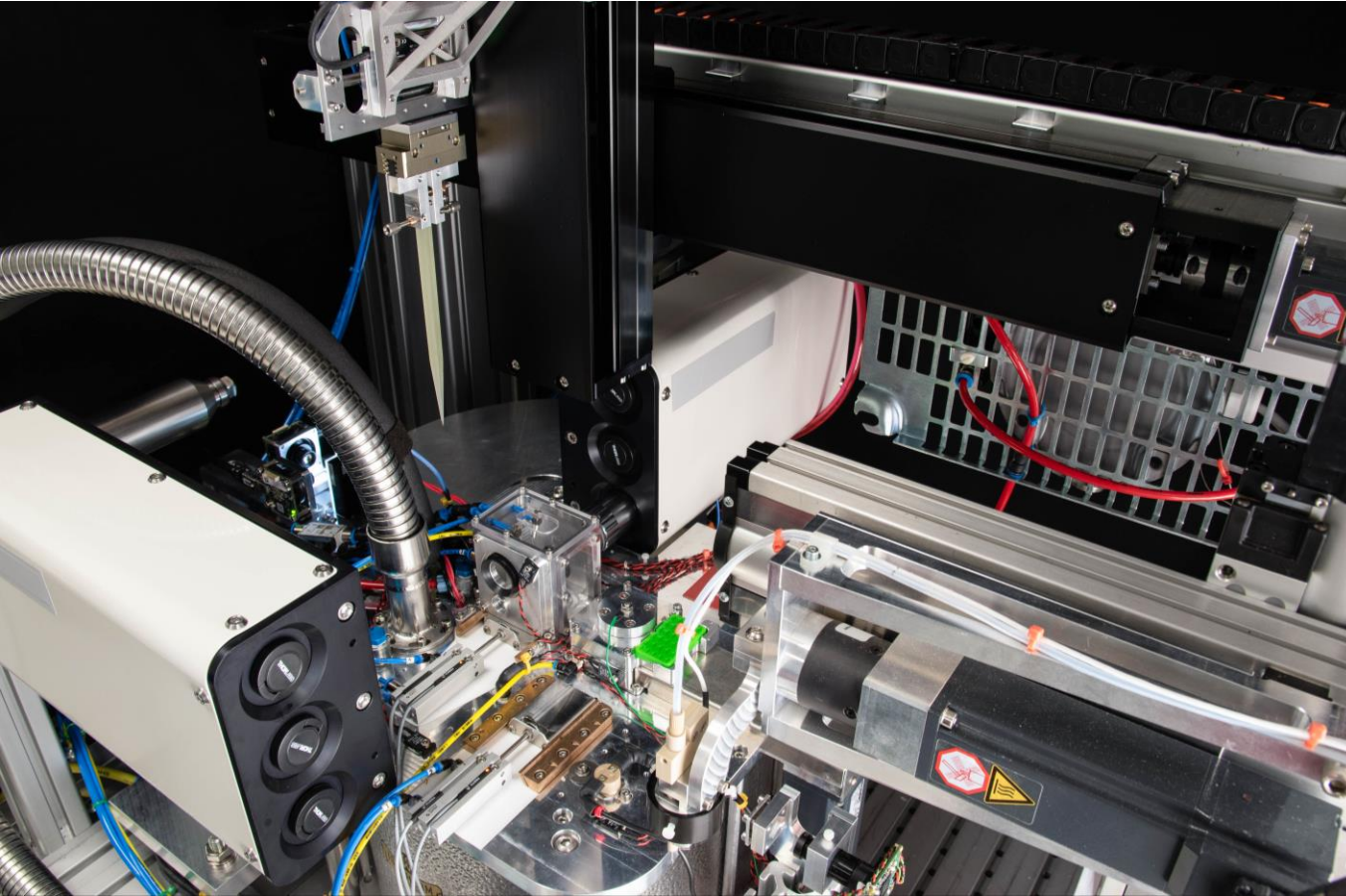
for vitrification

ographic

e (DHM)

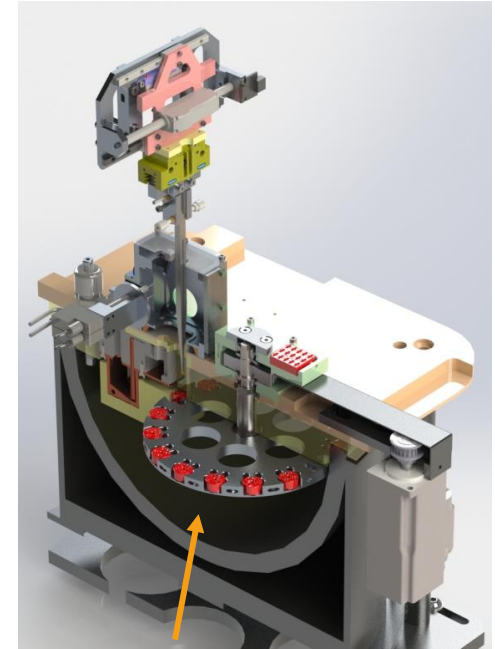
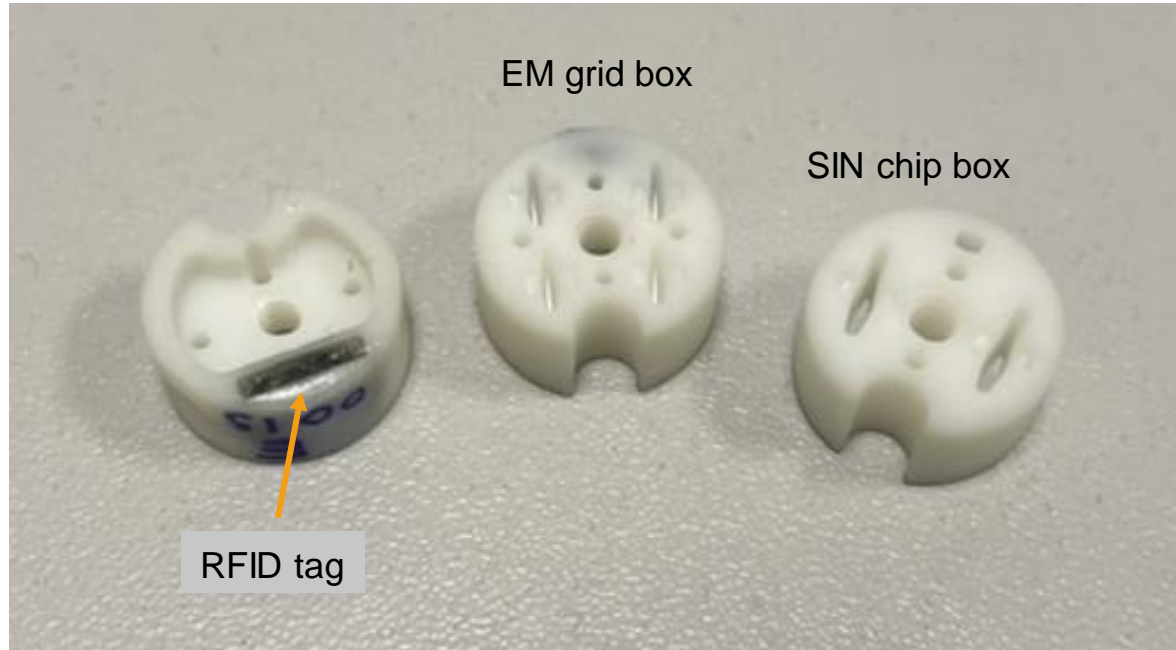
LN2

Components

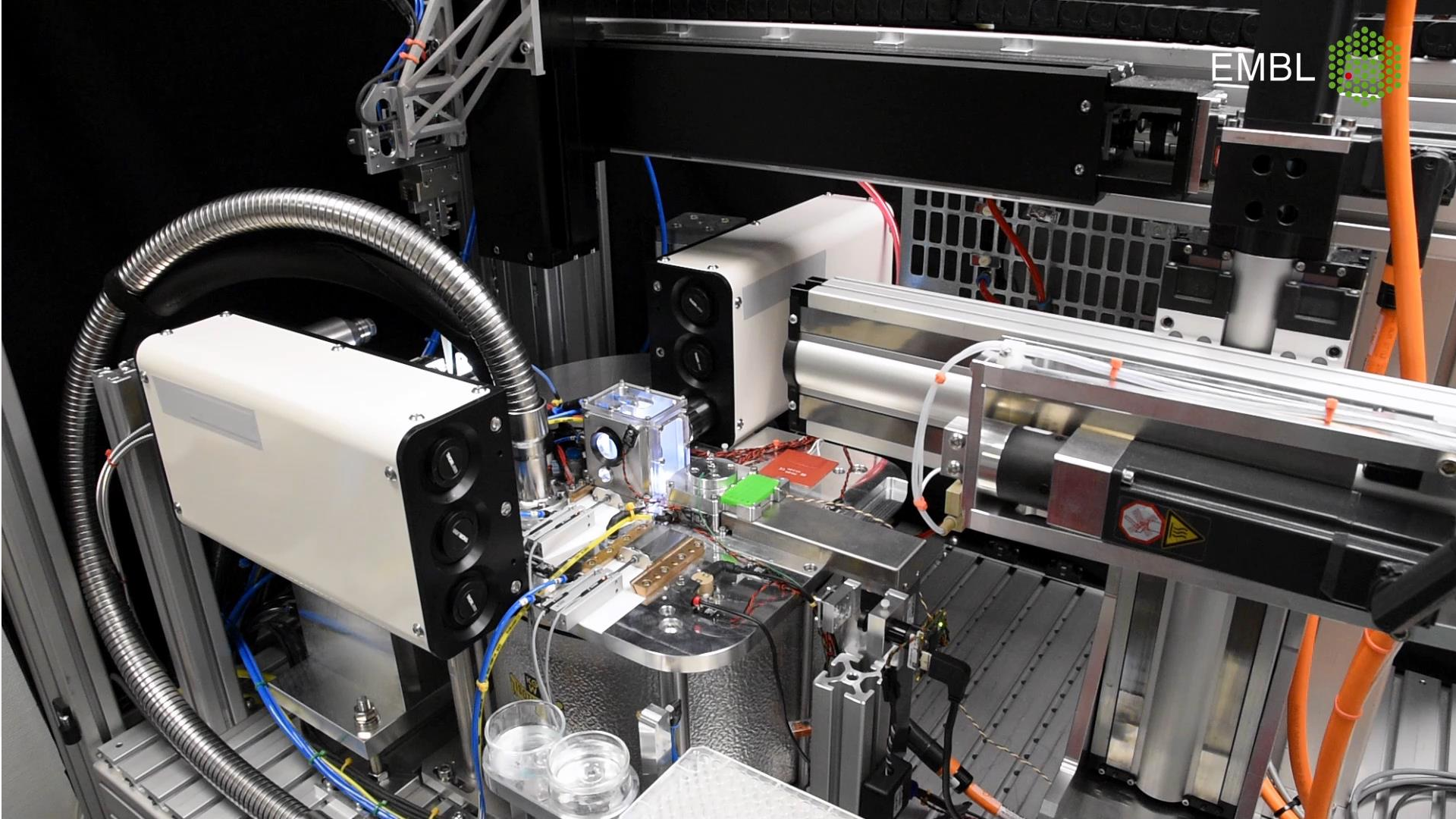


- Atmospheric plasma treatment
- Picoliter drop dispenser
- Chamber
 - Air blades (spreading)
 - Humidity control
 - Ethane jet for vitrification
- Digital Holographic Microscope (DHM)
- Dewar

The EasyGrid machine – sample storage



Capacity : 10 boxes = 40 EM grids



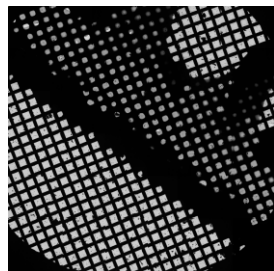
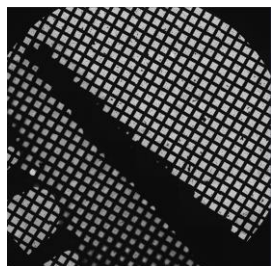
GUI

The screenshot displays the 'EasyGrid CryoEM sample preparation machine' GUI. The interface is organized into several functional areas:

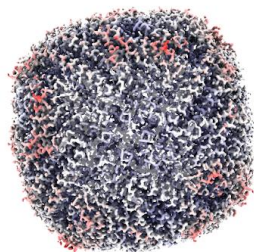
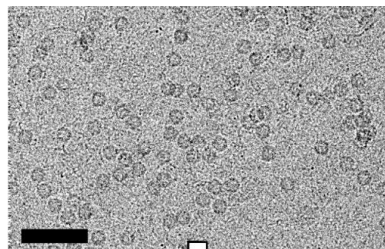
- Hardware Status (Top Left):** Shows system health with 'OK' status and warnings for Explosive Gas (Off) and Temperature Control (Enabled). It lists various temperatures: Target (20 °C), Solution Plate (23.4 °C), Pipettes (27.0 °C), Chamber (23.9 °C), Relative Humidity (16.23 %), and Dewpoint (-2.99 °C).
- Sample Loading / Sample Check (Top Center):** Features a 12x8 grid of wells. Blue wells represent 'Protein wells' and orange wells represent 'Coating wells'. Below the grid are buttons for 'Fill Protein', 'Fill Coating', 'Fill Both', and 'Clean'. A large circular image shows a sample with concentric rings.
- Dewar (Bottom Left):** Includes 'Cool Down' and 'Heat Up' buttons, an 'Abort (NI)' button, and 'Park Pipette Robot' and 'Park Gripper Robot' buttons.
- SPA grid preparation (Bottom Center-Left):** Shows 'Sample support Selection' with a grid of 5x6 wells. A yellow highlight is on the 3rd row, 4th column. 'Take' and 'Put back' buttons are present. A note states: 'NB : Grid goes to gripper check after "Take"'. Below this is a 'Plasma Treatment' section with a 'Plasma Treat' button and 'Nb passes: 10'.
- Dispensing parameters (Bottom Center-Right):** Includes 'Mode' (Static/Dynamic), 'Horizontal step (mm): 0.3', 'Points per line: 10', 'Burst frequency (Hz): 100', 'Line number: 1', 'Vertical step (mm): 0.8', 'Vertical offset (mm): 0', and 'Drops per burst: 10'.
- Blowing (Bottom Right):** Features 'Enable Blowing' (checked), 'Blowing Duration (ms): 30', and 'Delay before freezing (ms): 0'. There is also a 'Flash' section with 'Enable Flash' and 'Last Flash DeltaT: 0.0 ms'.
- Sample Selection (Bottom Far Right):** A circular dial with 12 positions (1-12) and 'Empty' options. It includes 'Type: None', 'Code', and '# Code St...' fields. 'Coating' and 'Prepare' buttons are located below the dial.

At the bottom left, the status bar shows 'Ready Admin 16:50:27'.

Purified protein structures



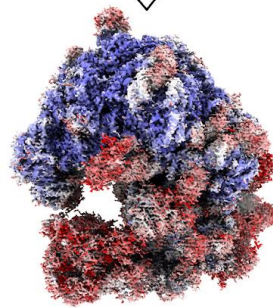
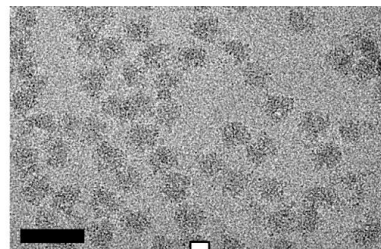
apoferritin



1.92 2.02 2.12
Local resolution (Å)

$R_{avg} = 1.9\text{Å}$

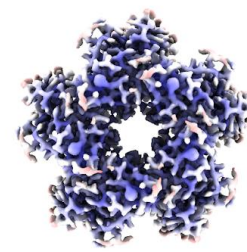
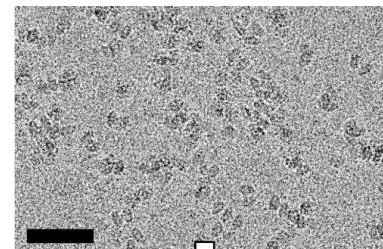
ribosome



1 5 9
Local resolution (Å)

$R_{avg} = 2.4\text{Å}$

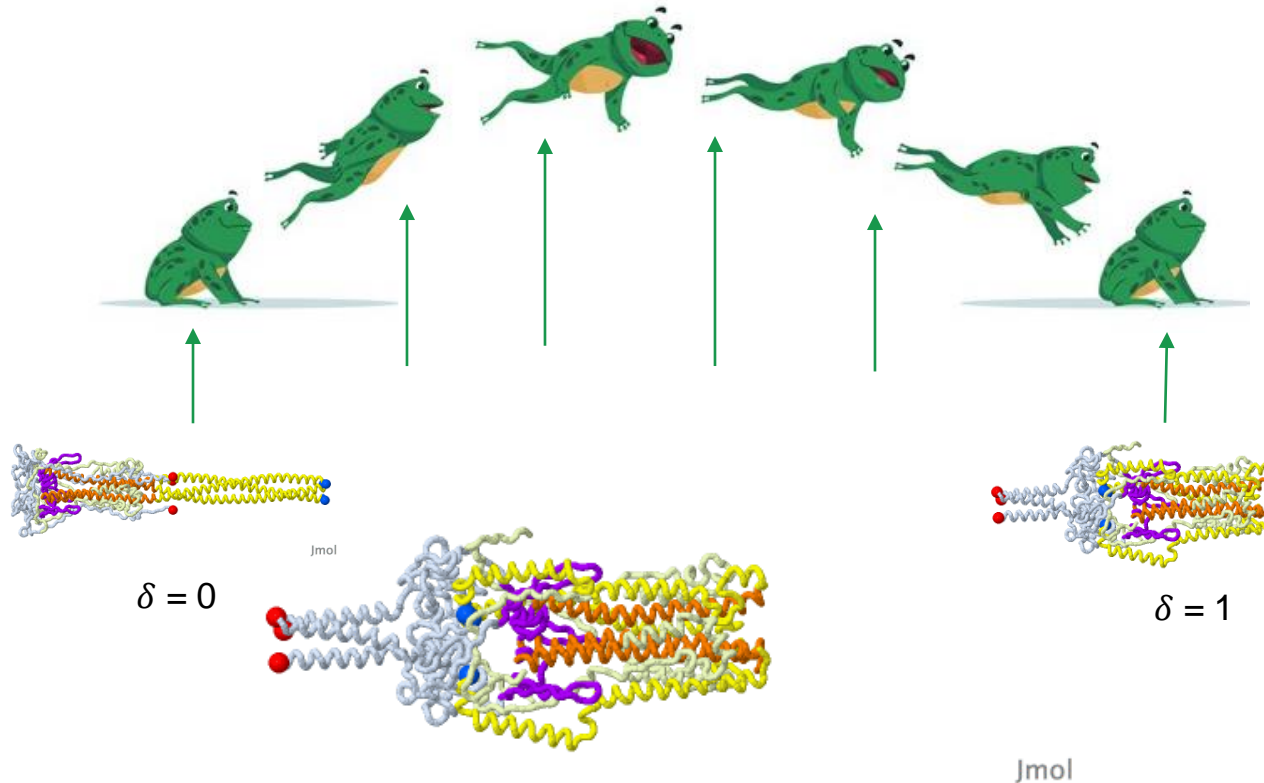
rhodopsin



2.10 2.45 2.80
Local resolution (Å)

$R_{avg} = 2.3\text{Å}$

Time-Resolved experiments - Towards movies of biochemical reactions



adapted from protopedia.org

Time-Resolved experiments - Towards movies of biochemical reactions

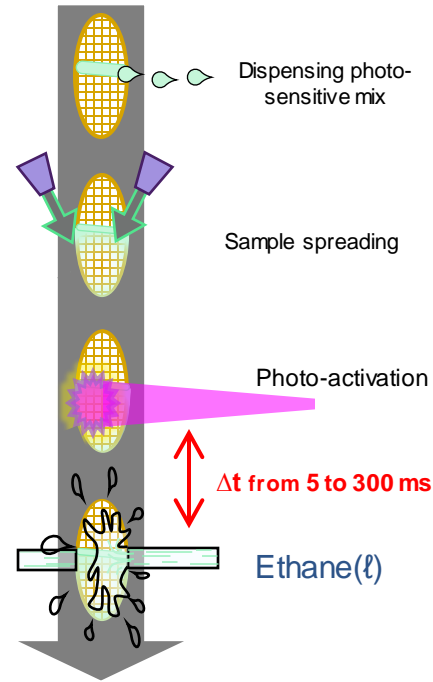
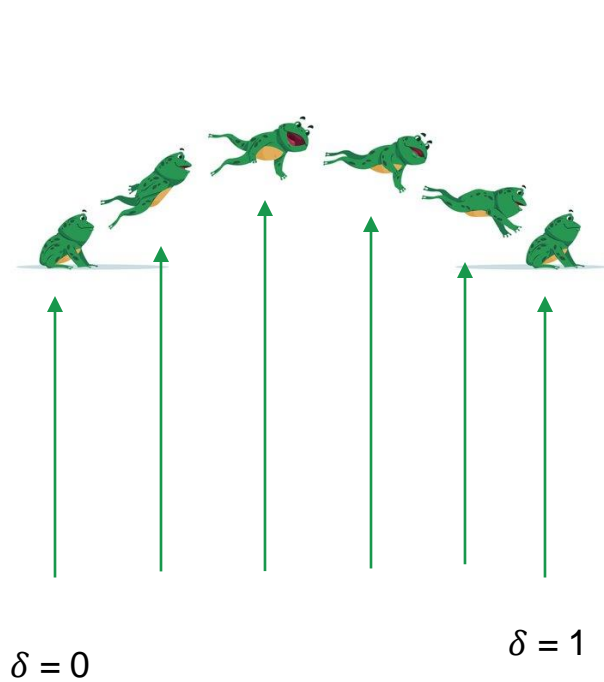
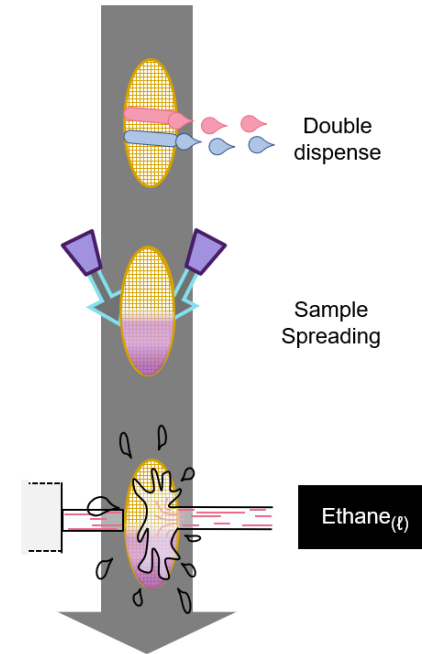
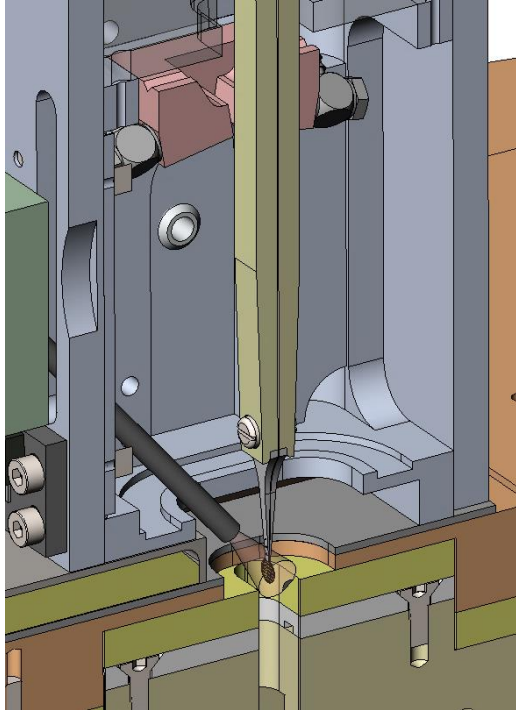


Photo-activation
module

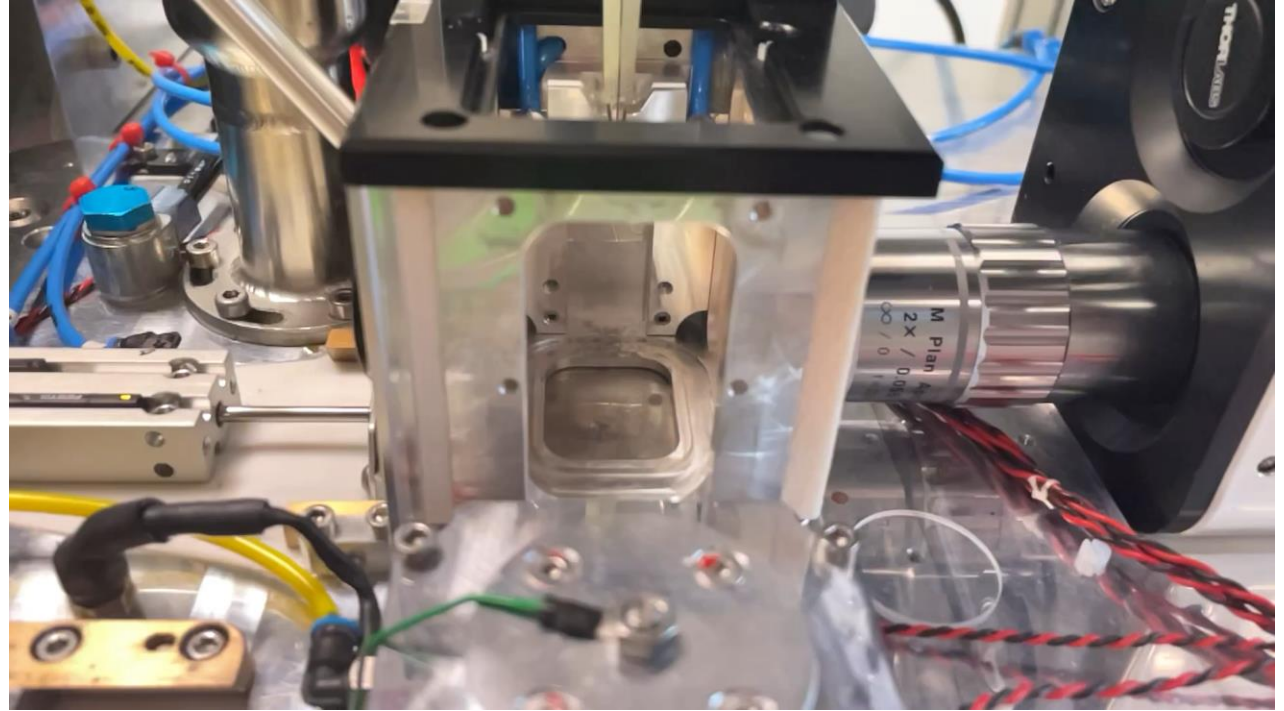


Mixing with double
dispense

Towards time resolved studies (light triggerig)



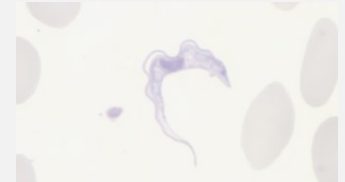
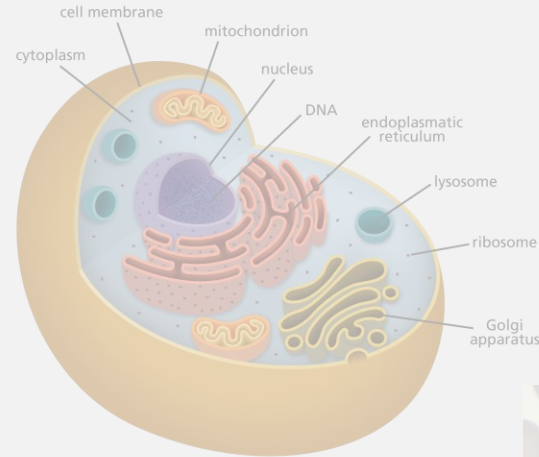
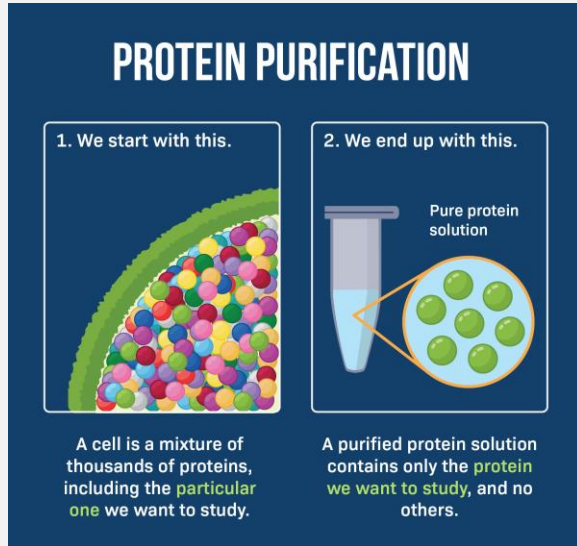
Light triggering setup (CAD)



Light triggering video – **Slow motion**

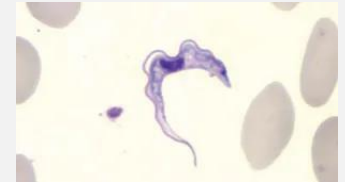
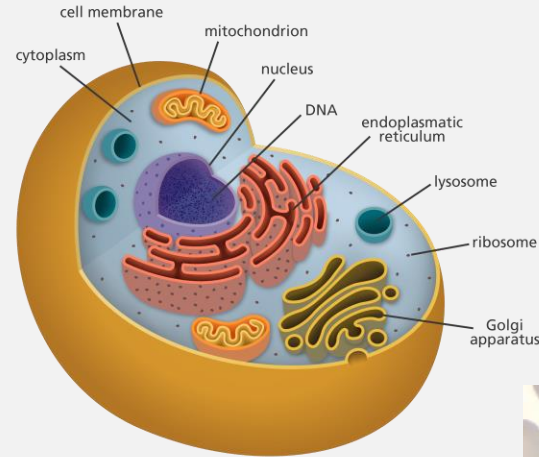
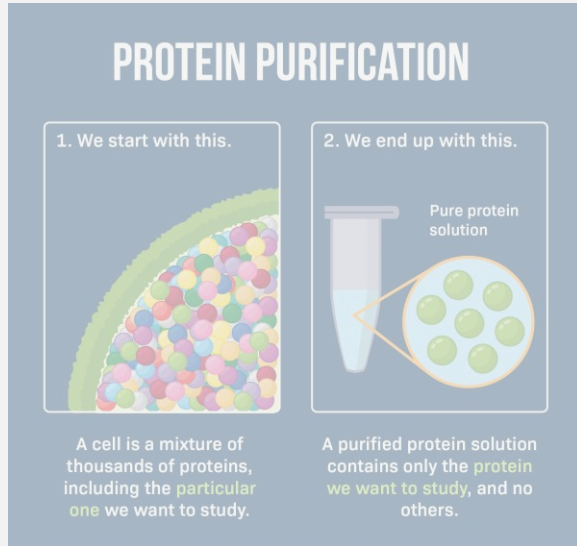
Which samples?

- Purified protein solutions (Single Particle Analysis)
 - Structure determination at atomic resolution
- Cells, monocellular organisms
 - In-situ study, proteins in their natural environment

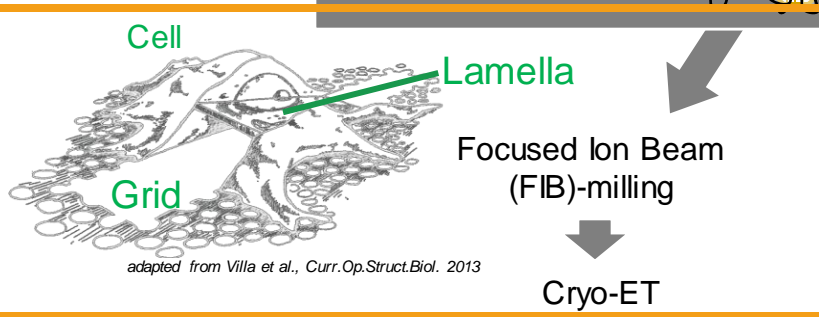
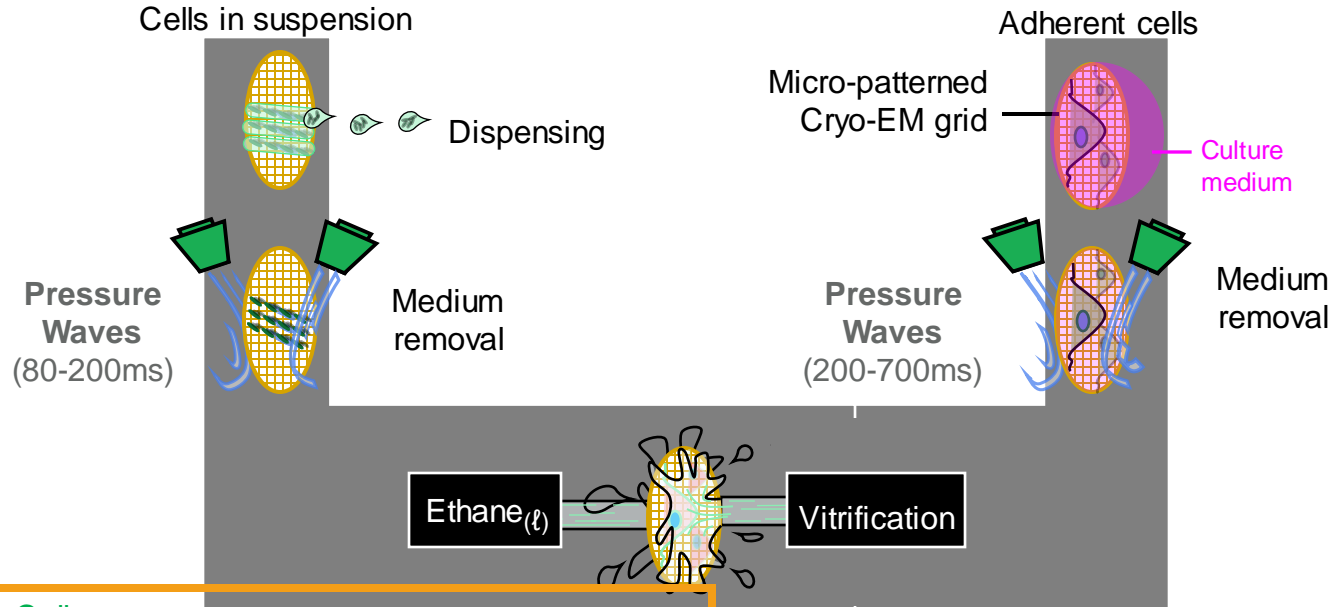


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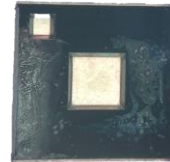
- Purified protein solutions (Single Particle Analysis)
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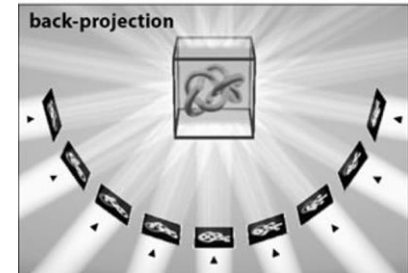
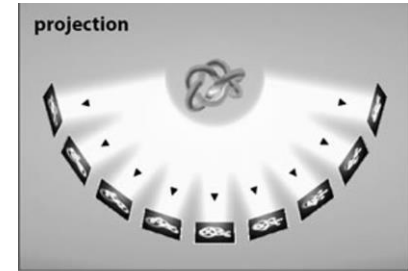
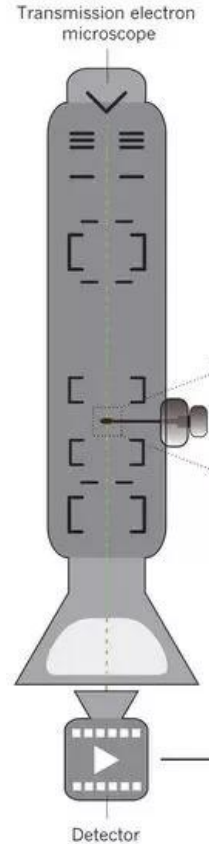
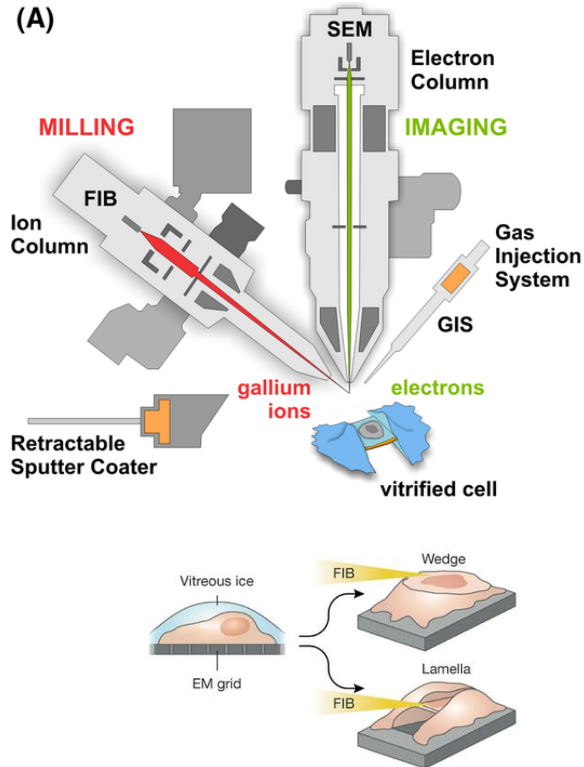
Cell vitrification for in-situ imaging



X-Ray imaging



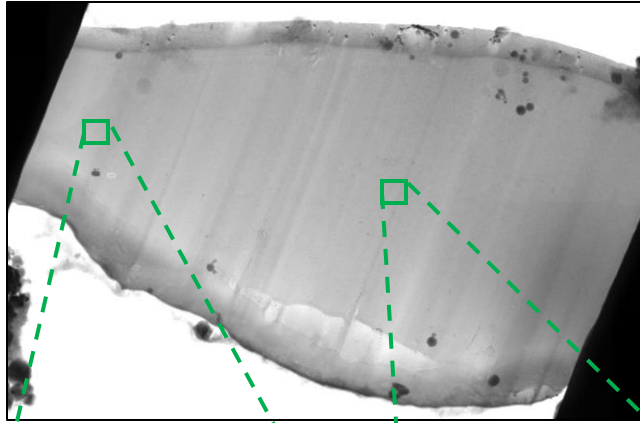
Cryo-ET FIB-SEM and data collection



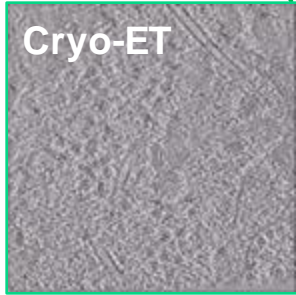
Cryo-Electron Tomography of HeLa cells



Lamella of *jet*-frozen HeLa cell (EasyGrid)



Cryo-ET

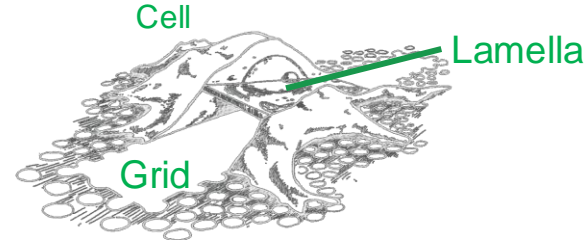


Cytosol

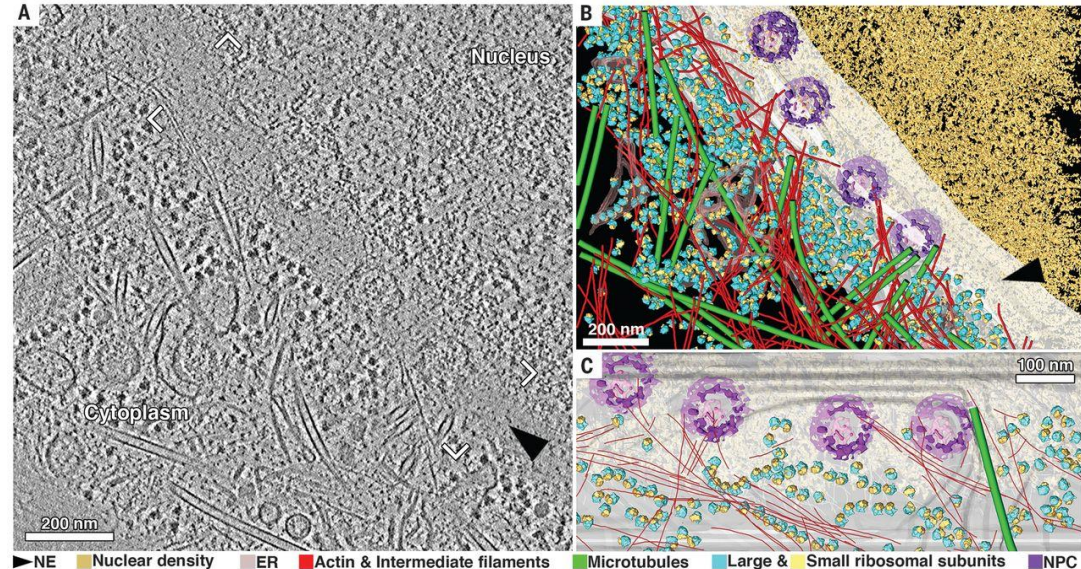
Cryo-ET



Nucleus



adapted from Villa et al., *Curr.Op.Struct.Biol.* 2013



Julia Mahamid *et al.*, Visualizing the molecular sociology at the HeLa cell nuclear periphery. *Science* 351, 969-972 (2016). DOI: [10.1126/science.aad8857](https://doi.org/10.1126/science.aad8857)

Acknowledgements

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 - Marcos Lopez-Marrero (alumnus)
 - Florent Cipriani (alumnus)
 - Kévin Lauzier (alumnus)
 - Christopher Rossi (alumnus)
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- Kowalinski Group:
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 - Georg Wolf
- EM support
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 - Sarah Schneider (alumna)
- Mattei Team
 - Olivier Gemin (alumnus)
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 - Zhengyi Yang
 - Georg Wolf
- Mahamid Group:
 - Steffen Klein
 - Anastasiia Babenko (alumna)
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- Schneider Group
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- IBS
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 - Felix Weis
 - Martin Weik
 - Benoit Gallet

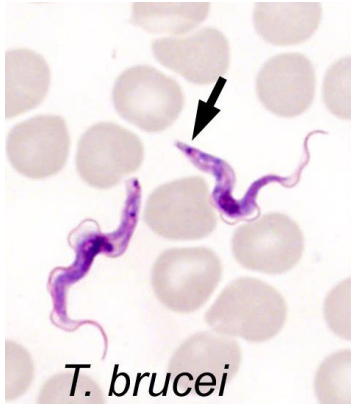


Thank you

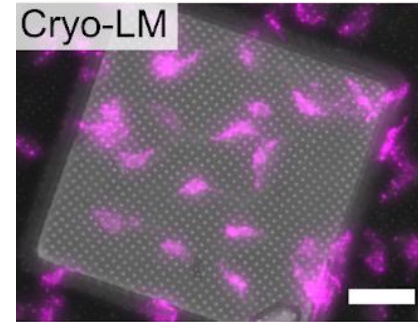
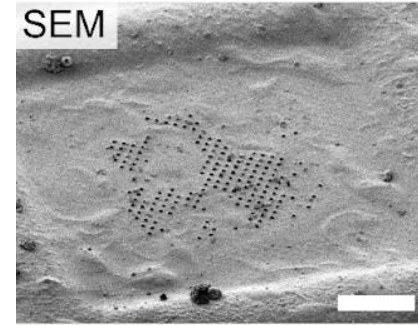
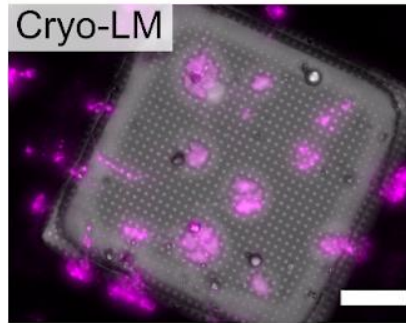
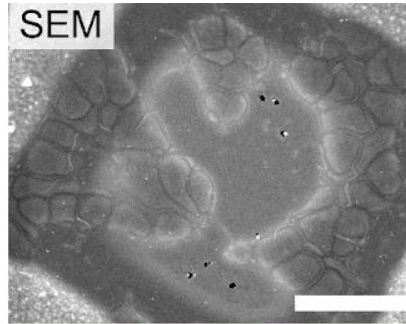
Questions?

Preserving natural shape (*Trypanosoma brucei*)

Leica GP2



EasyGrid



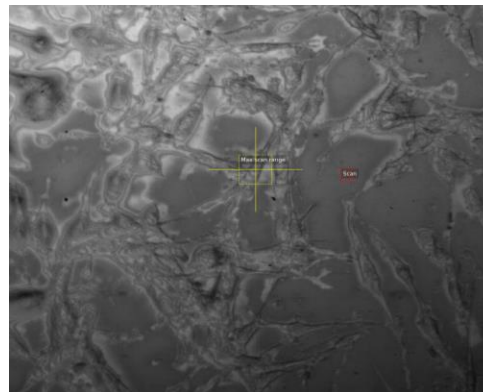
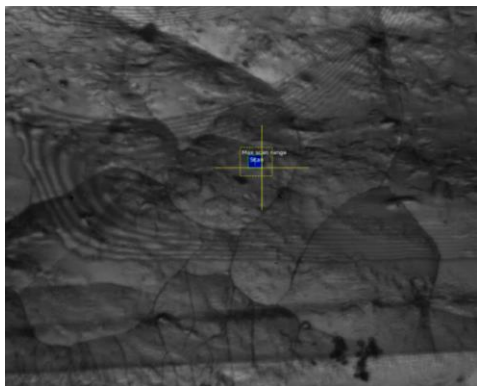
X-ray nano imaging (ESRF ID16A)

Toxoplasma Gondii infected HFF cells (24h post infection)

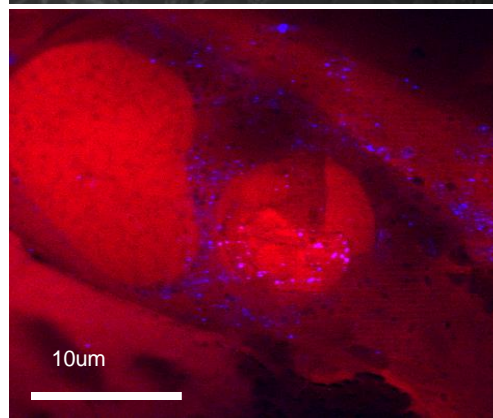
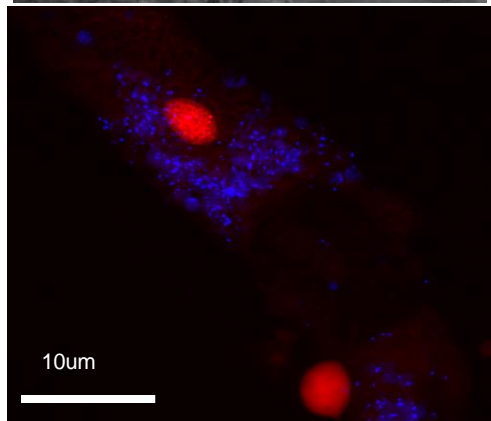
Leica cryo-plunger

EasyGrid

Phase images



XRF image
K/Fe
distribution map
(area density ng/mm²)



Cryo-Electro Tomography principle

Improvement in ice quality (HeLa cells)

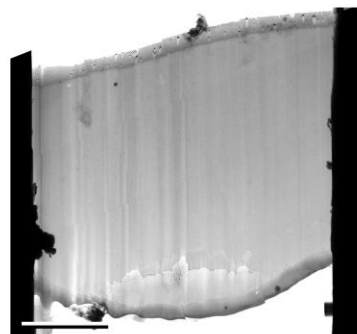
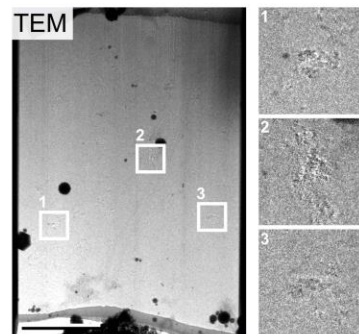


Leica GP2

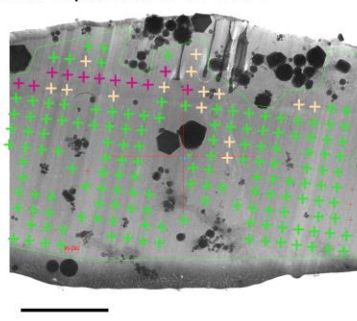
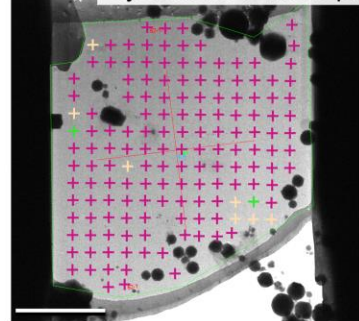
EasyGrid

a Plunge-frozen cells (N=20)

Jet-vitrified cells (N=16)



Systematic TEM acquisition at a pixel size of 1.2Å

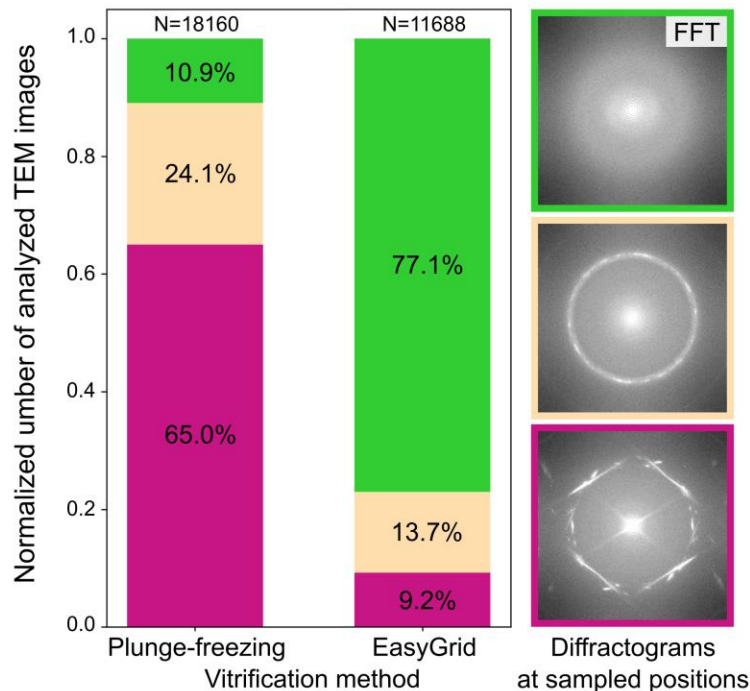


Vitreous

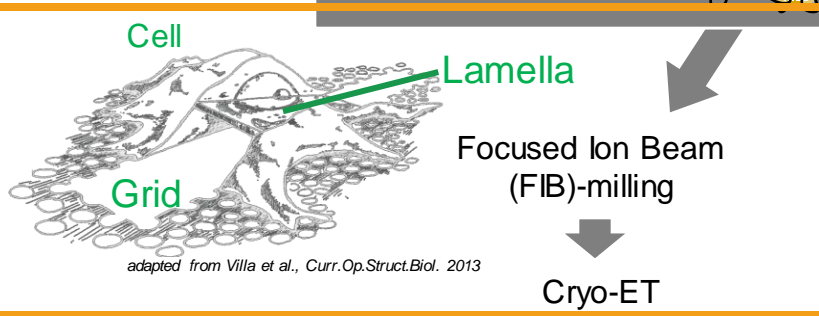
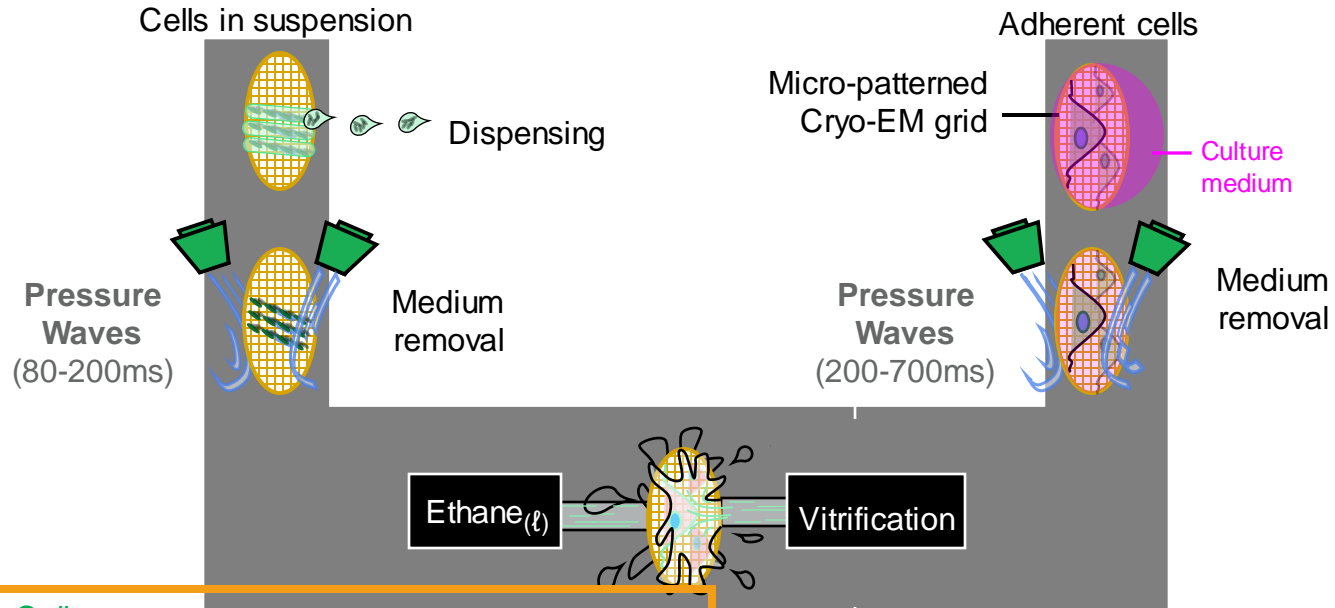
Semi-crystalline

Crystalline

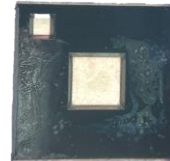
b Ice quality analysis using raster cryo-EM



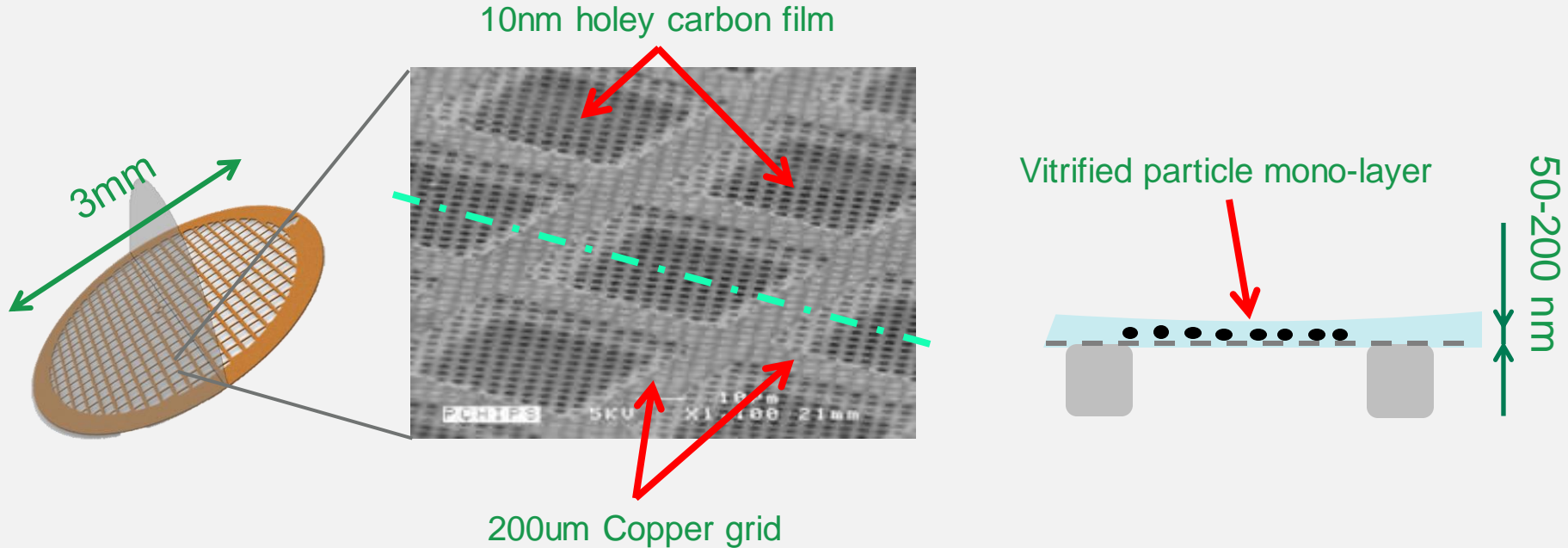
Cell vitrification for in-situ imaging



X-Ray imaging

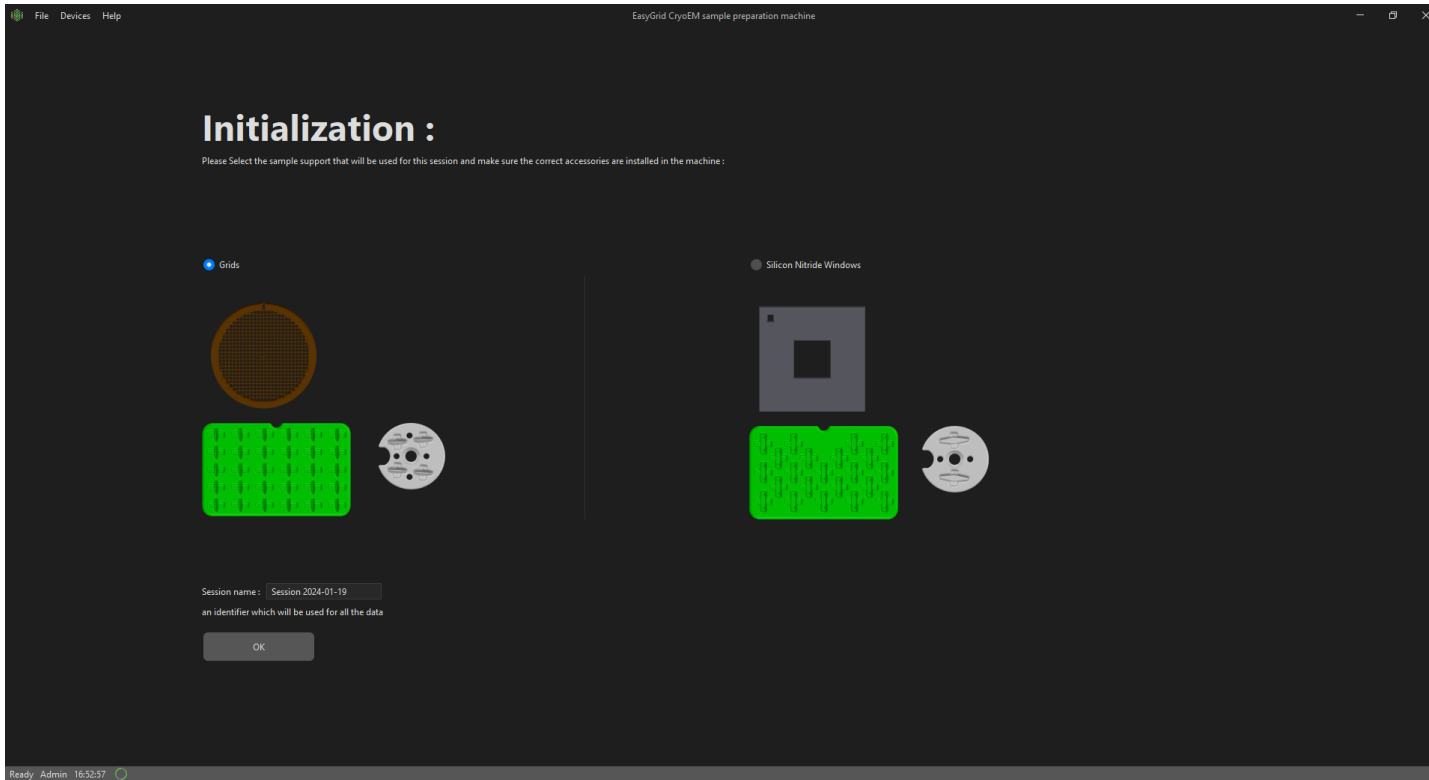


Ideal sample for Single Particle Analysis



Vitrification: rapid freezing of the sample to form amorphous ice and not crystalline.

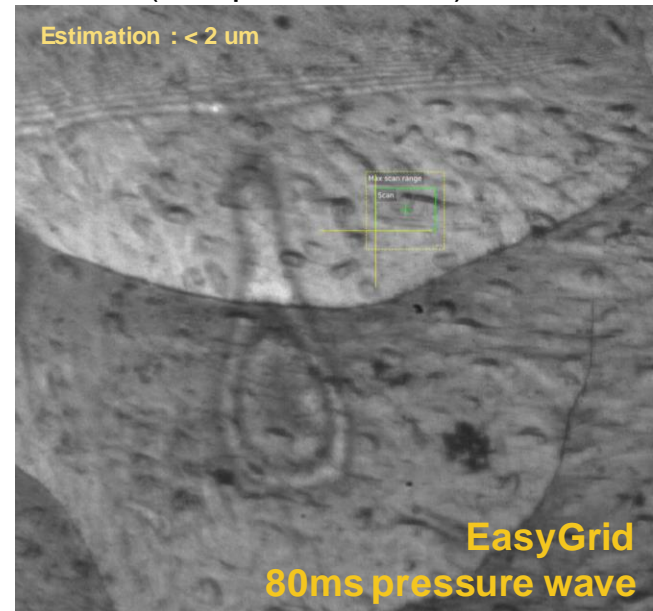
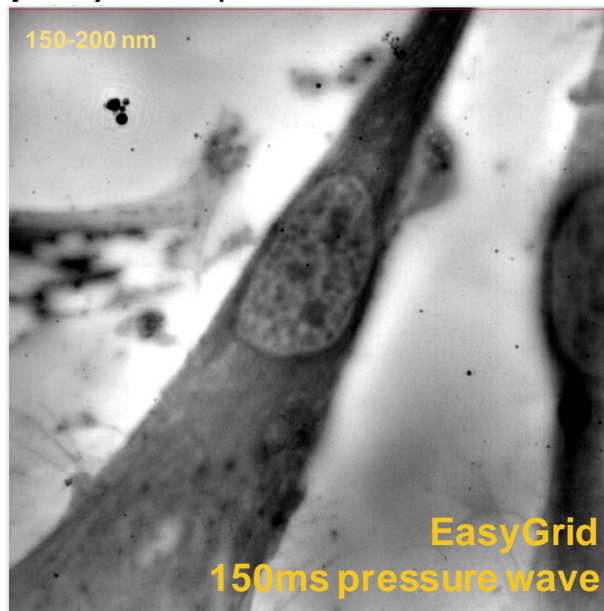
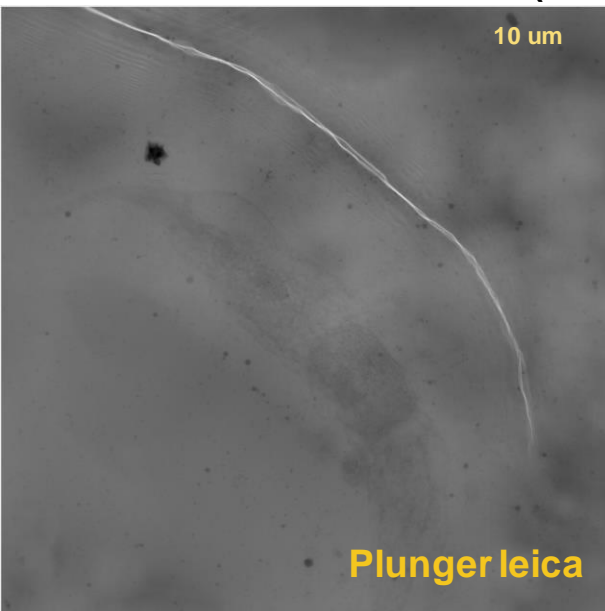
Rapid switch between sample supports



X-ray nano imaging (ESRF ID16A)

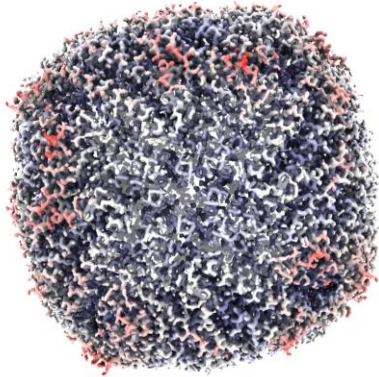


2D Phase Reconstructions (25nm/pixel) - Toxoplasma Gondii infected HFF cells (24h post infection)

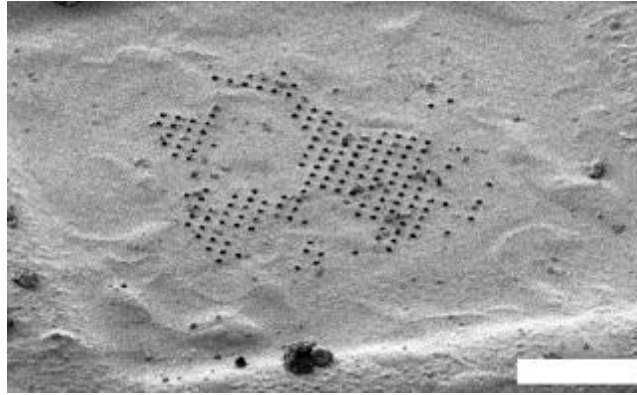


Ice layer thickness can be tuned

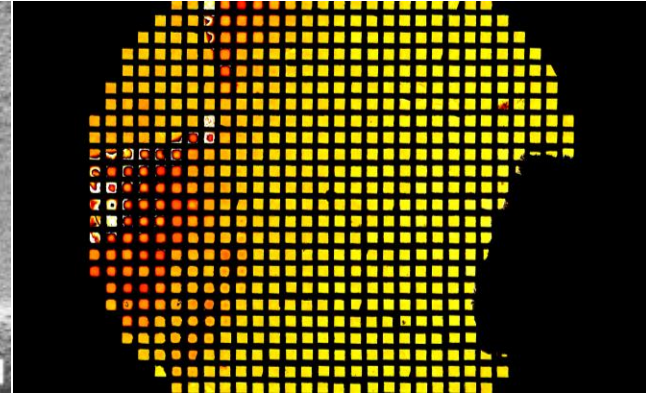
EasyGrid – use cases



Single Particle Analysis

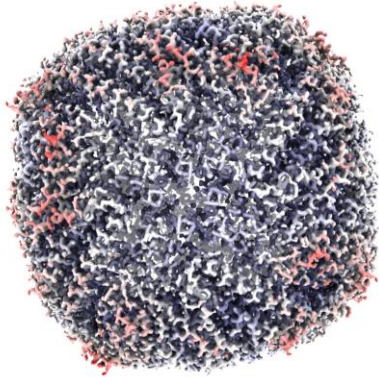


Cell vitrification

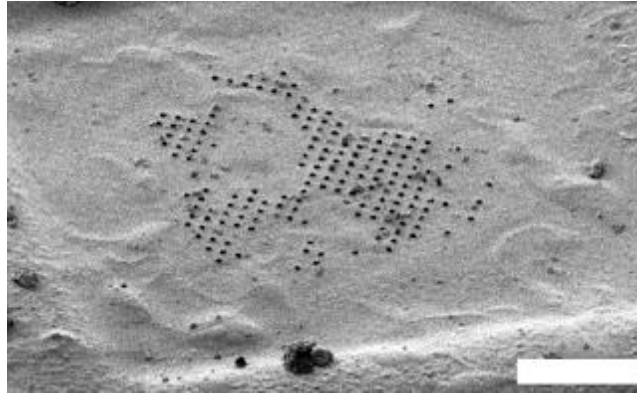


Quality control

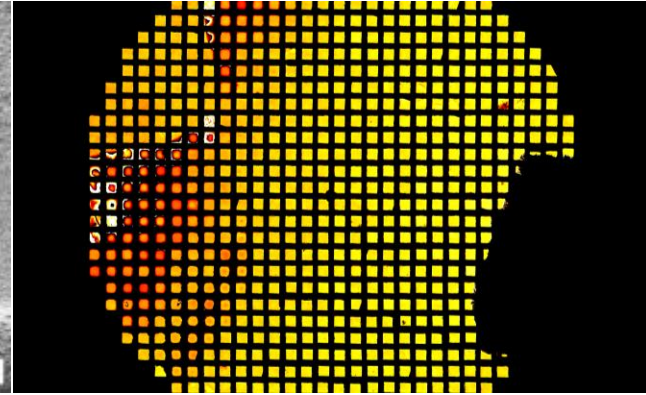
EasyGrid – use cases



Single Particle Analysis

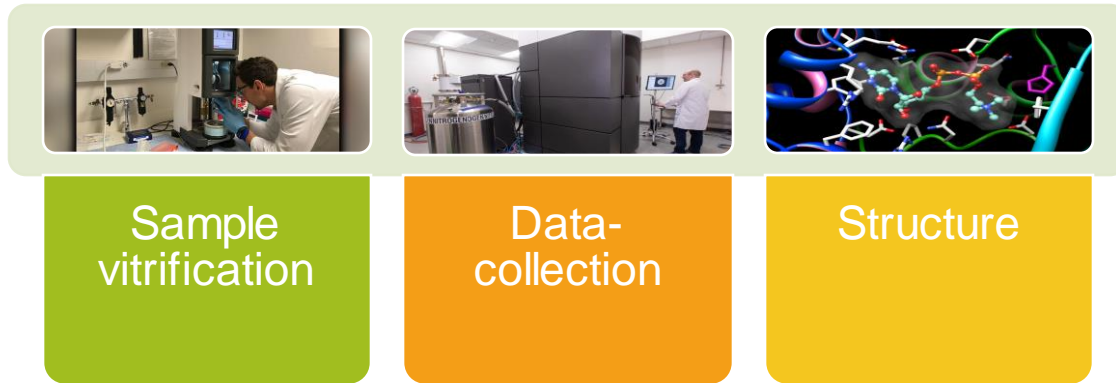


Cell vitrification

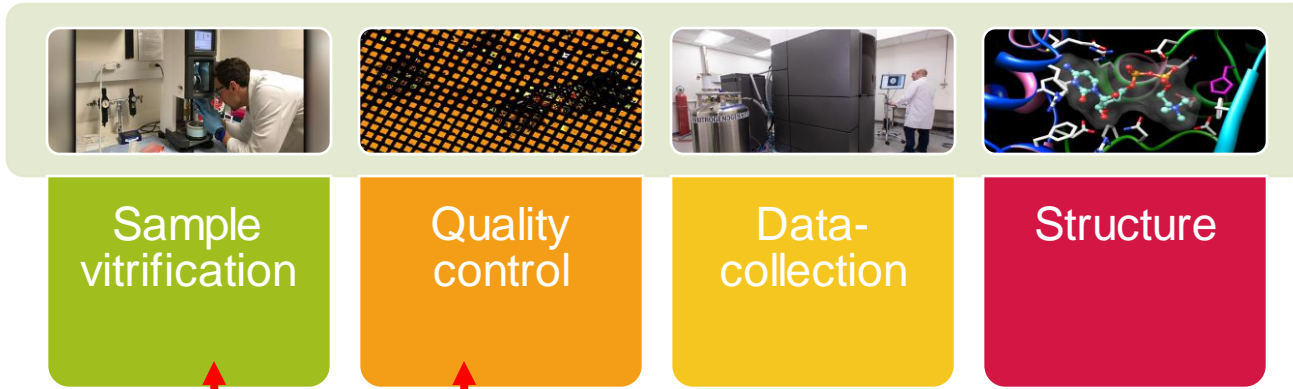


Quality control

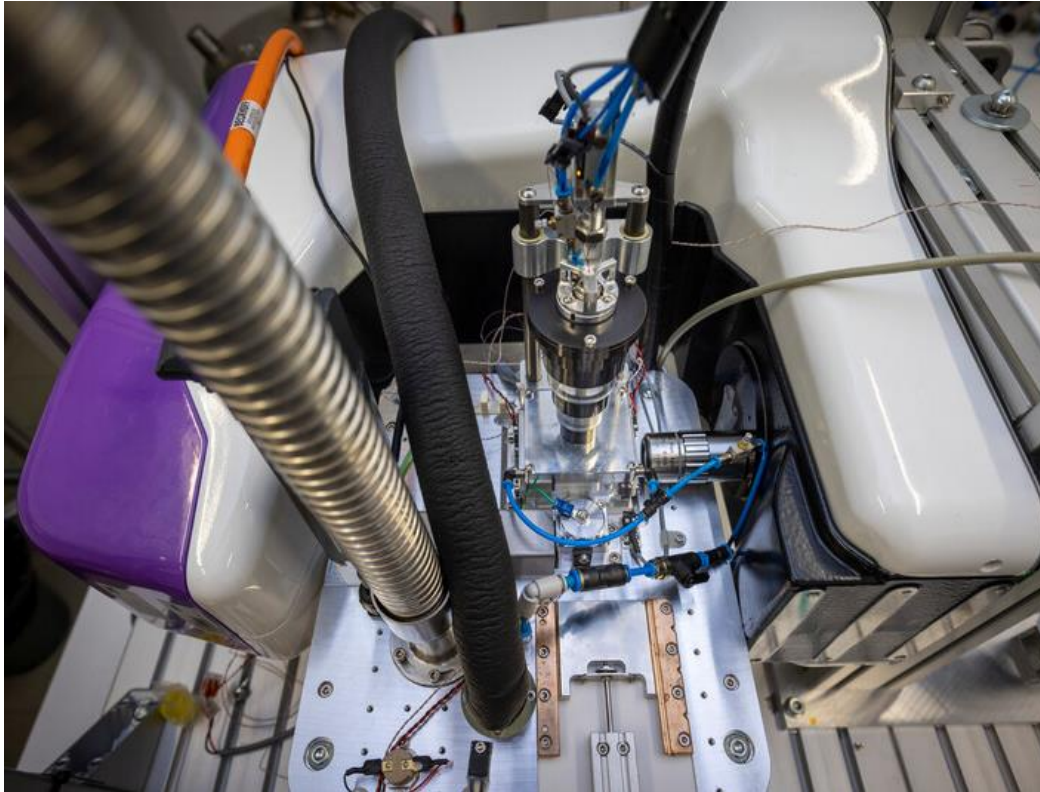
The traditional workflow for Cryo-EM SPA



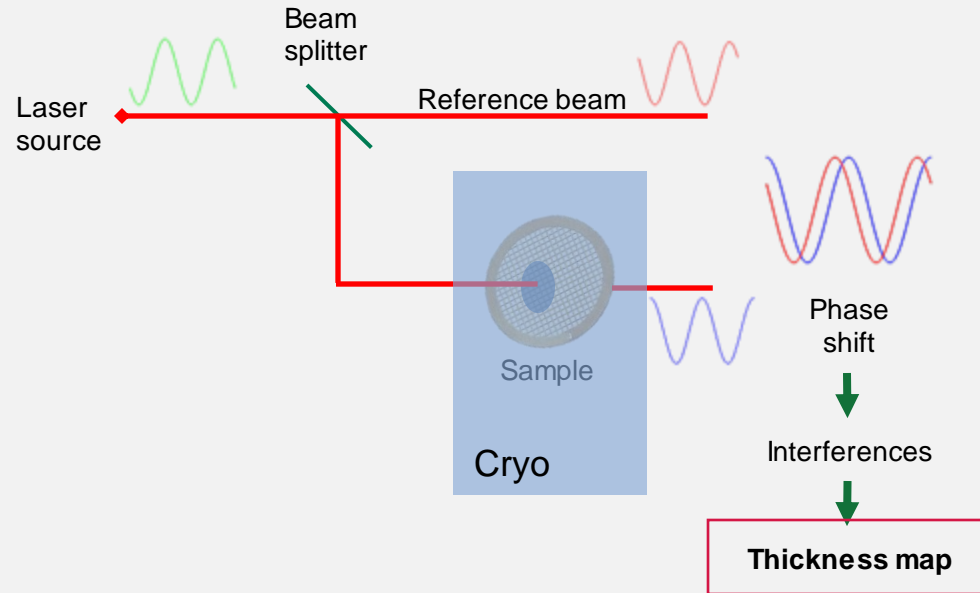
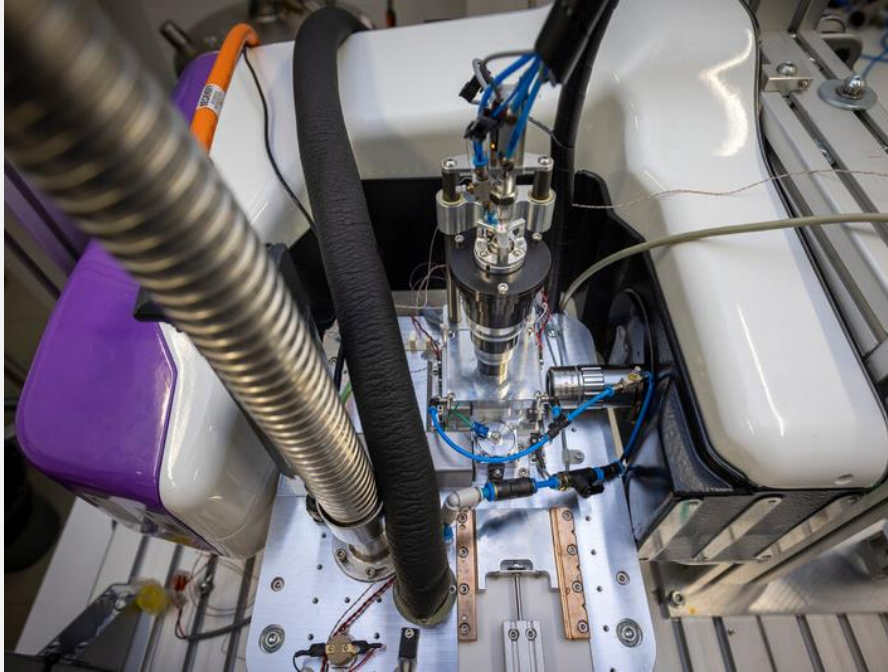
The EasyGrid workflow for Cryo-EM SPA



EasyGrid Control machine

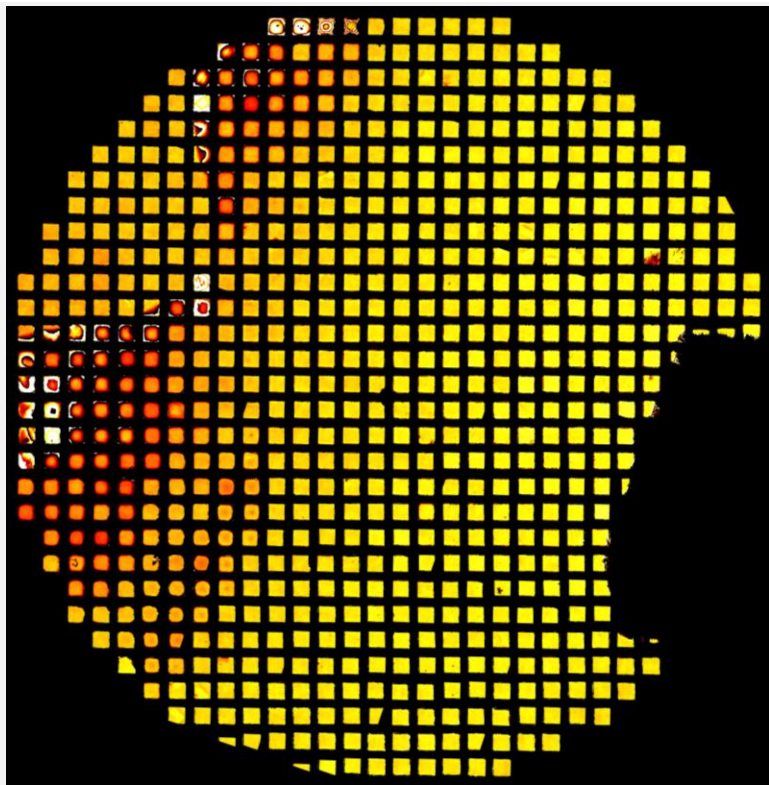


EasyGrid Control - principle



EasyGrid Control cryo observation column

SPA Grids – thickness map



EGC

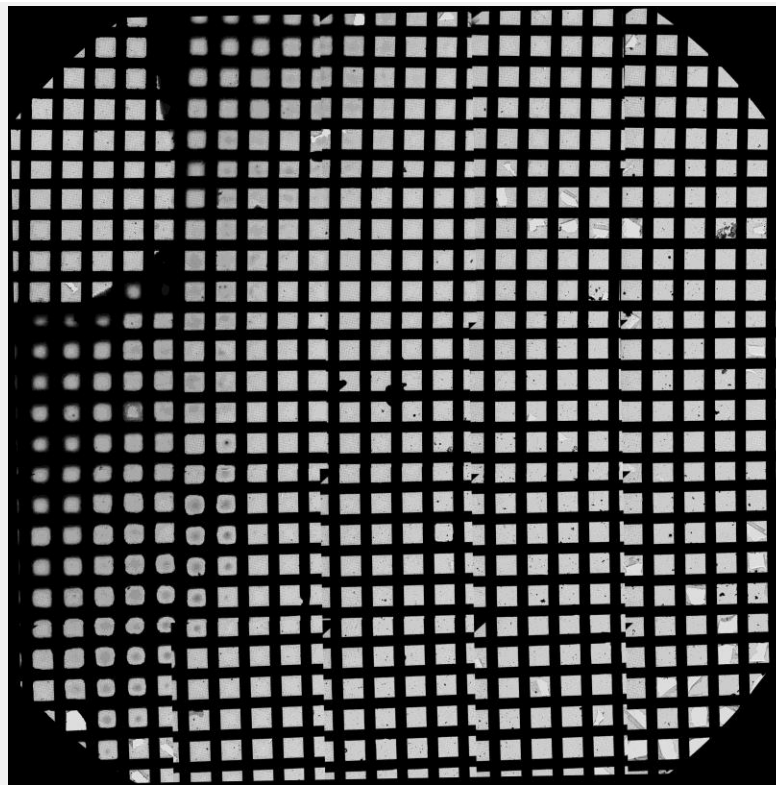
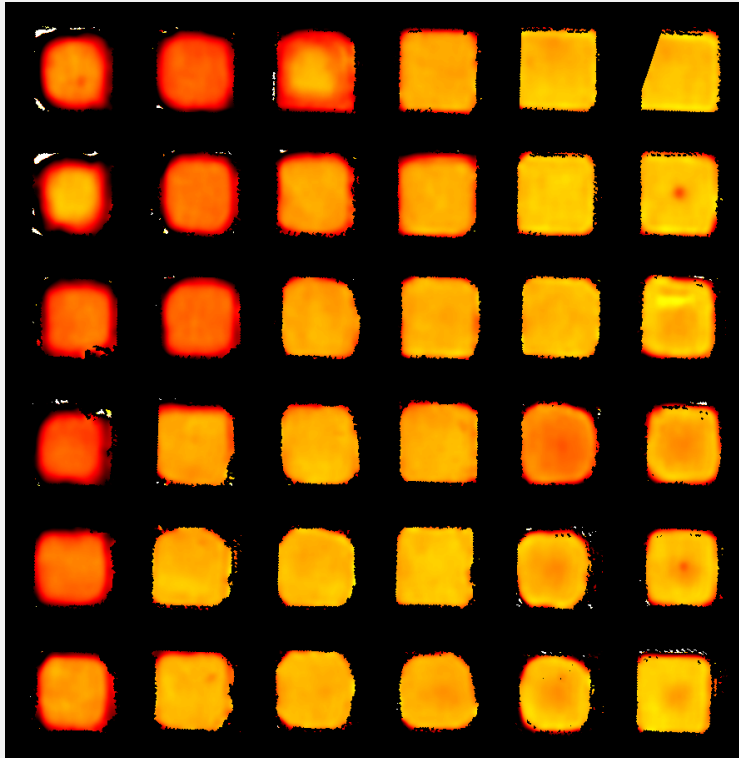
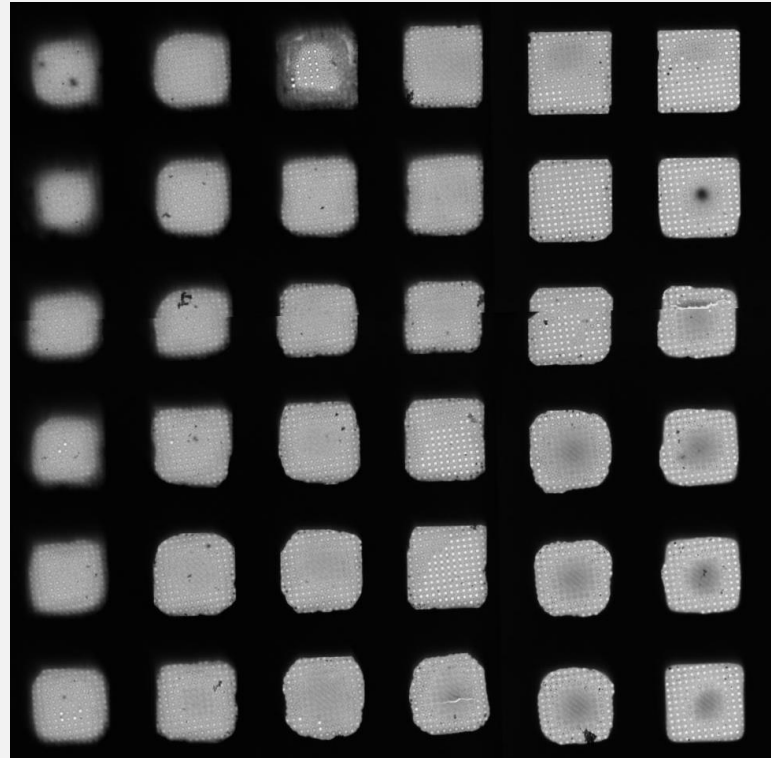


Image from Cryo-EM
Low resolution atlas

SPA Grids – thickness map



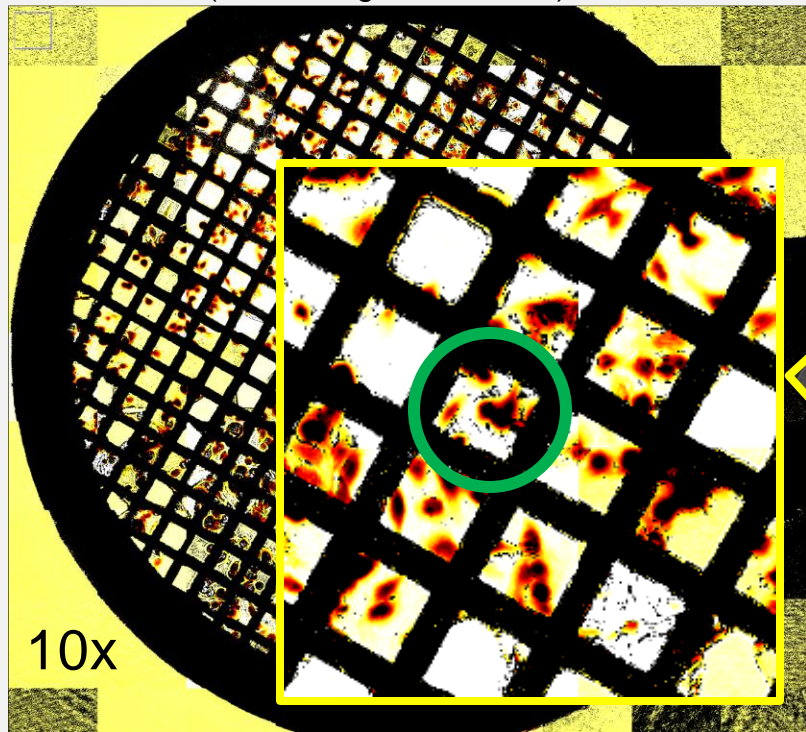
EGC



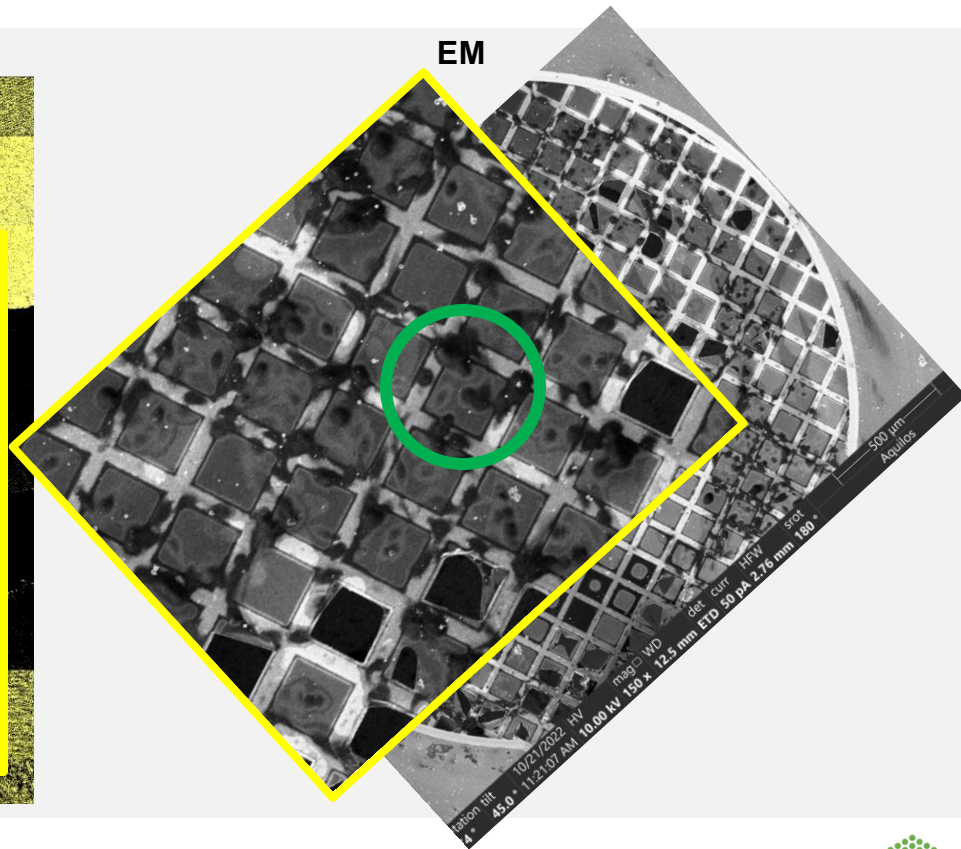
Glacios atlas

Grids with vitrified cells

EGC (HeLa2-4_g1 : BF650ms)



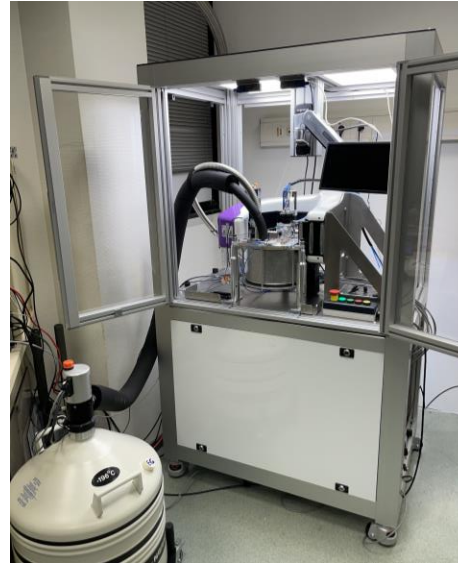
EM



Optimizing sample preparation



EasyGrid



EasyGrid Control



FIB-SEM



Cryo-EM



X-ray nano imaging

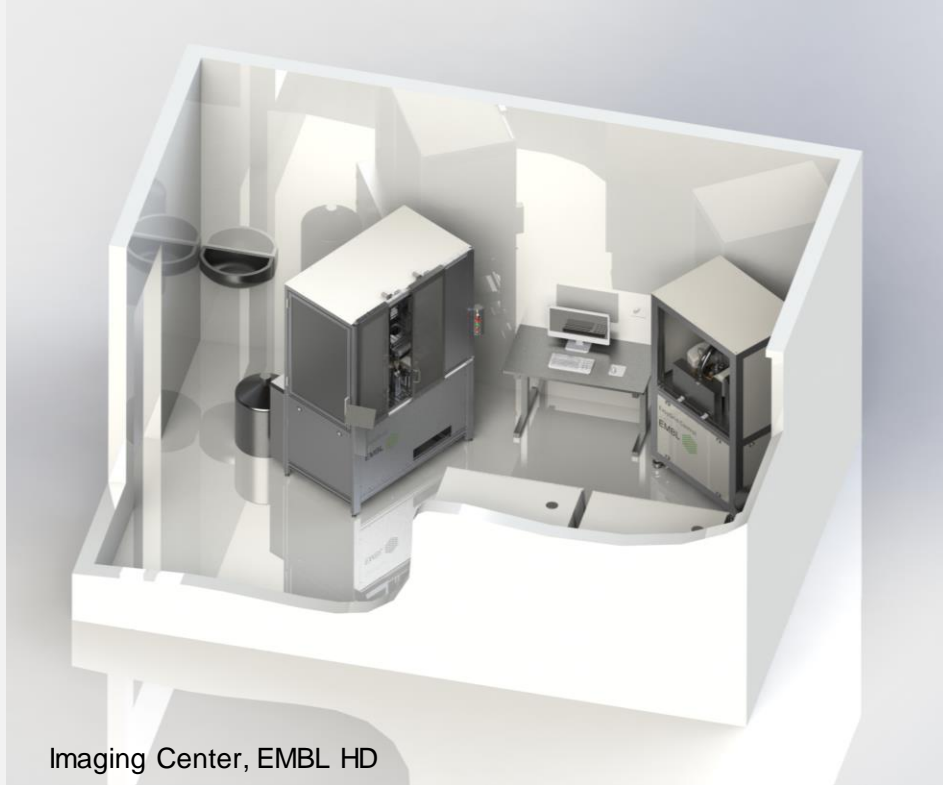
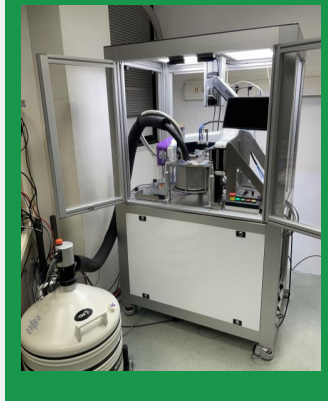
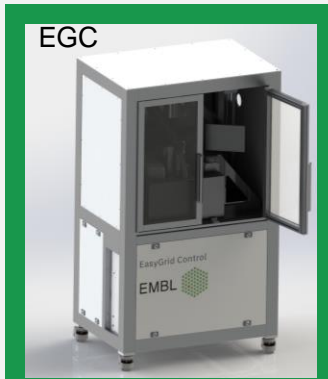
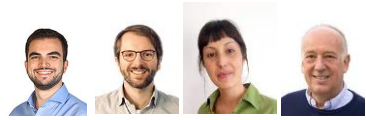
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Questions?

EasyGrid at EMBL-HD Imaging Center (2024)



Next steps

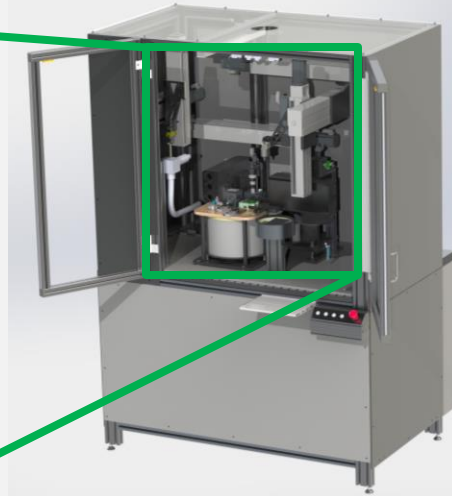
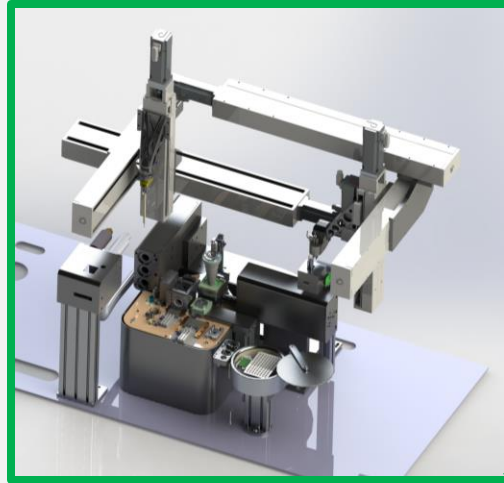
EasyGrid 2 (EG2)
Sample preparation

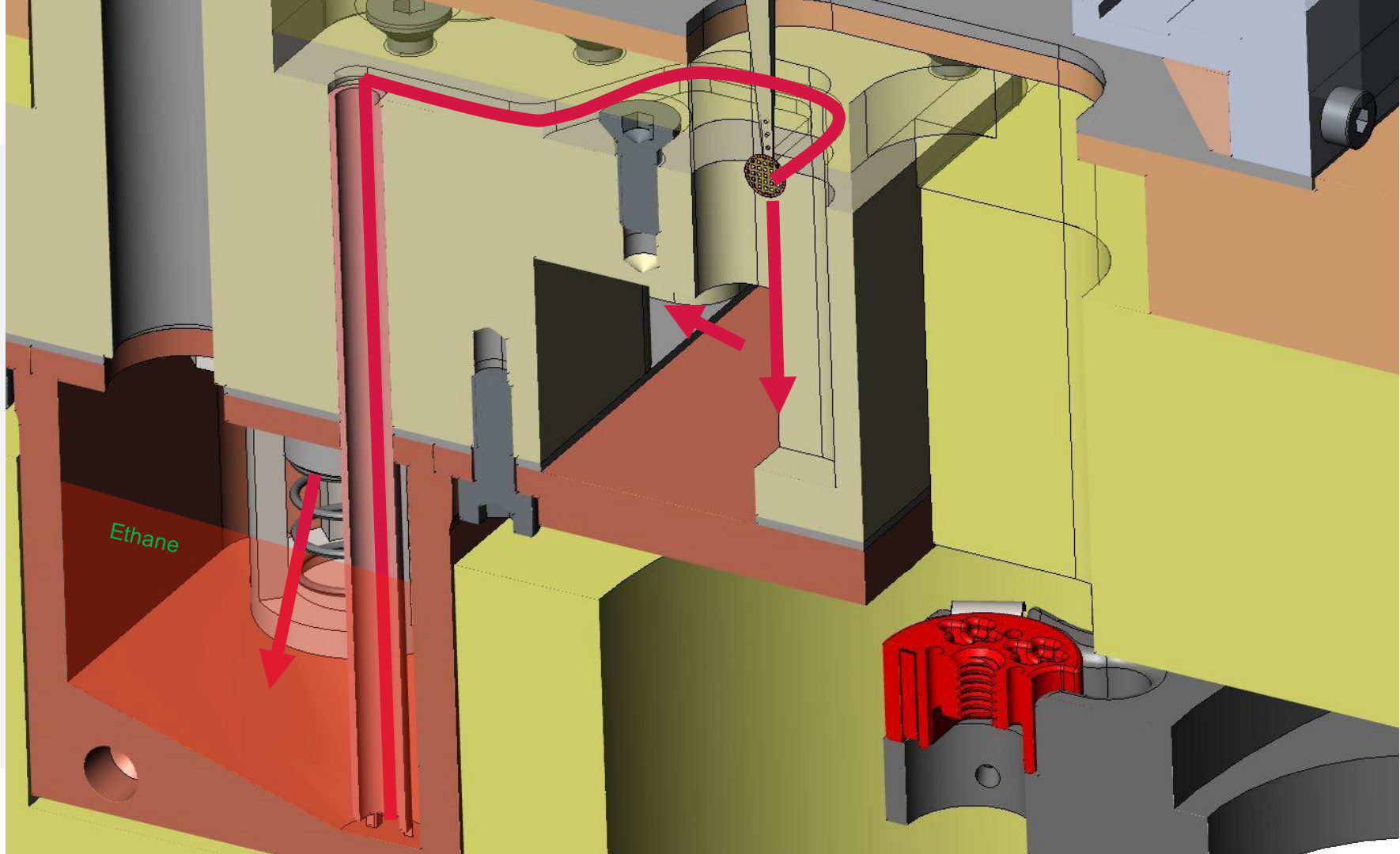
EasyGrid Control (EGC)
Sample control

Installation to EMBL-HD Imaging Center

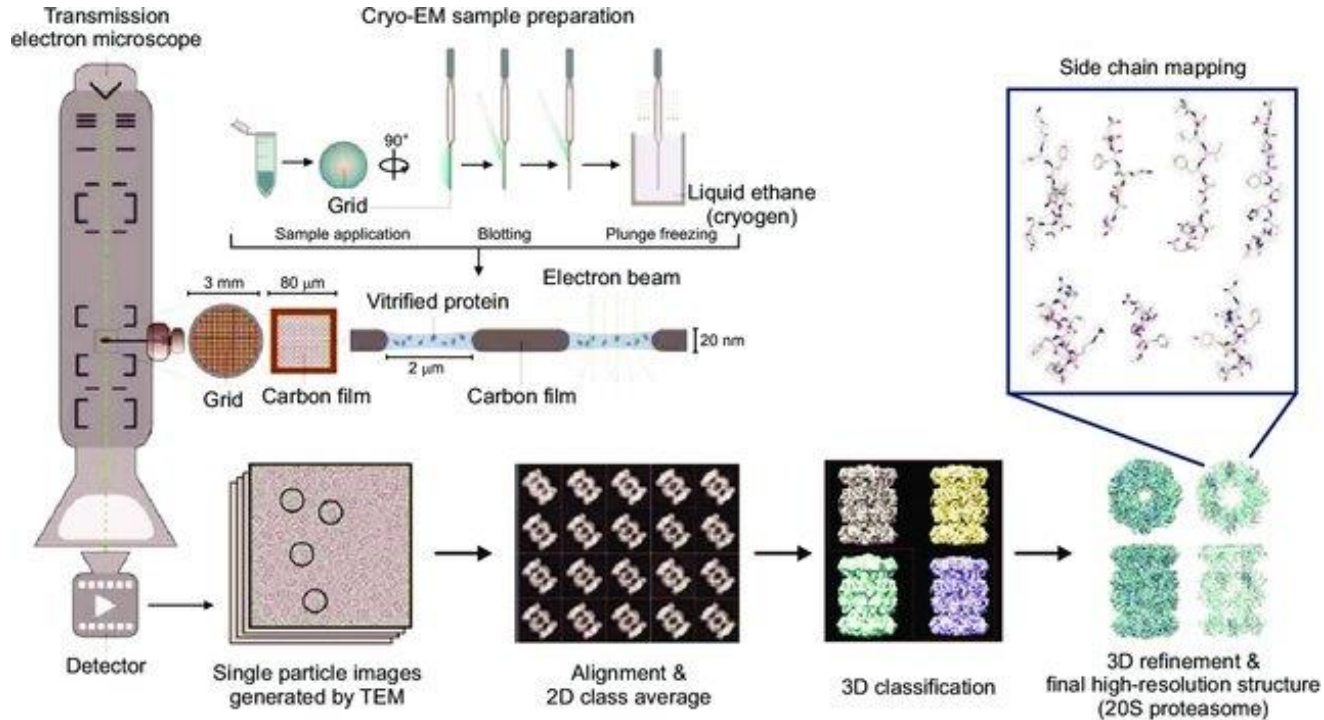
EasyGrid 3 (EG3)

Preparation & Control
Dedicated L2 lab @ EMBL-GR
Service provision - 2024





Cryo-EM principle



Determination of high-resolution structure through image processing